

Total Synthesis and Revision of Absolute Stereochemistry of Antillatoxin, an Ichthyotoxic Cyclic Lipopeptide from Marine Cyanobacterium Lyngbya majuscula

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Abstract—Antillatoxin is an ichthyotoxic cyclic lipopeptide isolated by Gerwick and co-workers from the marine cyanobacterium *Lyngbya majuscula* collected in Curaçao. Although we have finished the stereoselective total synthesis of antillatoxin having the proposed structure with (4S,5R)-configuration, we have found that the synthetic sample was not identical with the natural one and the proposed structure should be revised. Further our synthetic efforts have culminated in the first total synthesis of antillatoxin in its natural form, proving that the natural one has (4R,5R)-configuration. © 2000 Elsevier Science Ltd. All rights reserved.

Introduction

Marine cyanobacteria have proven to be exceptionally rich sources of structurally unique and biologically active natural products. Recently, Gerwick et al. reported their discovery and structural description of three distinct classes of structurally novel natural products from the marine cyanobacterium, *Lyngbya majuscula*, collected in Curaçao (Fig. 1). Curacin A was highly toxic to brine shrimp and also exhibited an antiproliferative activity due to its inhibition of tubulin polymerization.¹ Further studies of curacin A have established that it binds with high affinity to the colchicine site of tubulin and consequently inhibits the binding of colchicine.² This encouraging biological activity has motivated a significant world-wide effort towards the total synthesis of curacin A in a remarkably short time.³ Barbamide was isolated from the same cyanobacterium by using snail bioassay and showed a strongly molluscicidal activity.⁴ Its structure consists of a trichloromethyl portion and a thiazole amine portion, which appeared in dysidin⁵ and dolastatin **10**,⁶ respectively. In addition to curacin A and barbamide, a collection of *L. majuscula* furnished



Figure 1. Natural products from the marine cyanobacterium Lyngbya majuscula.

Keywords: antillatoxin; ichthyotoxicity; cyclic lipopeptide; total synthesis.

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the ichthyotoxic compound, antillatoxin.⁷ Therefore, Gerwick et al. have speculated that this array of bioactive metabolites functions in nature protects this cyanobacterium from predation by crustacea, herbivorous fish and gastropod mollusks, which are abundant in the marine habitat in which this organism thrives.

Antillatoxin (1) was isolated in low yield as an amorphous powder (1.3 mg, 0.07% of extract) from the ichthyotoxic crude extract of this marine cyanobacteria.⁷ The gross structure of 1 was established using extensive NMR studies including COSY, LR COSY, and HMBC. The absolute stereochemistries of antillatoxin-derived N-methylvaline and alanine were determined by acid hydrolysis of 1, HPLC separation and by chiral phase TLC versus standards. Both amino acids were present only as the l configuration. The stereochemistry at C4 and C5 was investigated using a combination of NOESY data, J values, CD spectroscopy and molecular modeling. Thus, the absolute stereochemistry at C4 and C5 was assigned as 4S.5R configuration, which was the lowest energy conformation in the modeling result. Antillatoxin is a structurally novel lipopeptide with a high degree of methylation. Especially, it has a conjugated diene that contains a tert-butyl group and a terminal olefin. In addition, a cis amide bond between alanine and N-methylvaline appears in the cyclic skeleton as a single rotational isomer. Goldfish toxicity measurements with antillatoxin showed it to be among the most ichthyotoxic metabolites isolated to date from a marine plant ($LD_{50}=0.05 \ \mu g \ ml^{-1}$), and it is exceeded in potency only by the brevetoxins (BTX-

Synthetic strategy

Our convergent synthetic strategy to (4S,5R)-antillatoxin (**1a**), having the proposed structure, is shown in Fig. 2. As we believed that the isolated terminal olefin of antillatoxin would be easily isomerized, we first employed the use of the phenylselenomethyl group as a precursor of this olefin and postponed the oxidative elimination of the phenylselenyl group to the final stages of the synthesis. Therefore, antillatoxin would be constructed from two subunits, a tripeptide unit **2** and a diene fragment **3**. Segment condensation of these fragments followed by final macrolactamization would give the desired macrocycle.

Synthesis of the tripeptide unit

Preparation of the tripeptide unit **2** of antillatoxin was achieved in a stepwise manner from glycine ethyl ester, as shown in Scheme 1. Coupling of (*S*)-Boc-*N*-methylvaline with glycine ethyl ester was carried out using diethyl phosphorocyanidate (DEPC, $(EtO)_2P(O)CN)^{12}$ to give the dipeptide **4** in 86% yield. After removal of the Boc group from **4** with trifluoroacetic acid (TFA), the amine was condensed using bis (2-oxo-3-oxazolidinyl)phosphonic chloride (BopCl)¹³ as the coupling reagent with allyloxycarbonyl (Alloc)-protected alanine in 50% yield. The resulting tripeptide **5** was saponified to furnish the tripeptide unit **2**, which was one partner for the upcoming fragment condensation.



Scheme 1.

A $LD_{50}=0.003 \ \mu g \ ml^{-1}$). Antillatoxin is a structurally and biologically attractive marine natural product. As a part of our program toward the synthesis of biologically active peptides from marine origin,⁸ we have embarked on the total synthesis of antillatoxin.⁹ In this paper, we wish to disclose the full details of our synthetic efforts toward antillatoxin, which includes a revision of the absolute chemistry at C4 and C5.^{10,11}

Synthesis of the conjugated diene portion

We initially attempted the iterative Horner–Emmons protocol for the construction of the conjugated diene of antillatoxin. Pivalaldehyde **6** reacted with the phosphonate **7** to give the *E* isomer of **8** as the major product (E/Z>20:1) in 59% yield. In this reaction, no E/Z selectively was found using ethoxy analog **7**' in place of **7**. The ester **8** was then



Figure 2. Retrosynthetic analysis of antillatoxin.



Scheme 2.

reduced with diisobutylaluminum hydride (DIBAL), and the allylic alcohol oxidized using chemical manganese dioxide $(CMD)^{14}$ to afford the corresponding aldehyde. A second Horner–Emmons condensation of this aldehyde with the phosphonate **7**' was carried out under the Masamune–Roush conditions¹⁵ to give the conjugated diene **9** in 38% yield. Reduction of the conjugated ester **9** using DIBAL was carried out again to produce the desired alcohol **10** quantitatively.

As an improved route to the diene portion, the Suzuki coupling approach¹⁶ was employed. This approach began from 4,4-dimethyl-2-pentyne (**11**),¹⁷ which underwent the hydroboration with catecholborane followed by hydrolysis to give the vinylboronic acid **12**. This intermediate **12** was subjected to the Suzuki coupling with the known iodoalkene **13**¹⁸ in the presence of a palladium catalyst, arsine ligand and cesium carbonate to afford the dienyl alcohol **10** quantitatively in a single operation (Scheme 2).

Synthesis of (4*S*,5*R*)-antillatoxin (proposed structure)

For construction of the *cis* configuration at C4, C5 in the proposed structure by Gerwick et al., a *syn* selective asymmetric aldol reaction by the methodology of Evans¹⁹ was employed, as shown in Scheme 3.

Oxidation of the allylic alcohol **10** using CMD yielded the aldehyde, which was added to the boron enolate derived from the carboximide **14** to afford the corresponding aldol adduct **15** in good yield. Removal of the chiral auxiliary from **15** with alkaline hydrogen peroxide followed by methylation of the resulting carboxylic acid produced the methyl ester **16** in 60% yield. Protection of the secondary hydroxyl group of **16** as the triethylsilyl (TES) derivative and reduction of the methyl ester with DIBAL provided the primary alcohol **17** in 98% yield. Alternatively, the known hydroxy ester **21** prepared by the method of Baker¹⁸ could also be transformed into the alcohol **17** in three steps. The





Scheme 4.

stereochemistry of the alcohol **17** prepared using the Evans aldol methodology was also confirmed by this alternative approach (Scheme 4).

The alcohol **17** was then oxidized with tetrapropylammonium perruthenate (TPAP)²⁰ and this followed by a Still– Horner olefination²¹ to afford the (*Z*)-ester **18** in 74% yield. Although the susceptibility of the dienyl hydroxyl moiety toward dehydration under the acidic condition was observed, careful treatment of **18** with TFA caused the cleavage of the secondary TES ether and the acid-catalyzed lactonization simultaneously to give the lactone **19** in moderate yield. Stereoselective introduction of the phenylselenomethyl group (as a precursor for the isolated terminal olefin) to the α , β -unsaturated lactone **19** was accomplished using PhSeCH₂Li in the presence of HMPA²² to provide the selenolactone **20** in 73% yield as a single isomer. The stereochemistry of **20** was assigned by NOE experiments, as shown in Scheme 3.

Saponification of the lactone ring in **20**, protection of the resulting carboxylic acid as the allyl ester, and segment condensation with the tripeptide unit **2** using 1-[3-(dimethylamino)propyl]-3-ethylcarbodiimide hydrochloride (EDCI·HCl) gave the ester **23** in 78% yield. Treatment of **23** with Pd(Ph₃P)₄ in the presence of morpholine caused simultaneous removal of the *C*-terminal allyl group and the *N*-terminal allyloxy carbonyl group. However, final

macrolactamization of the liberated amino acid **24** using several reagents led to decomposition of the starting material.²³ Accordingly, we attempted the macrolactamization after elimination of the phenylselenyl group. Oxidative elimination of the phenylselenyl group of **23** proceeded with NaIO₄ to provide the terminal olefin **25** in 93% yield. After removal of the protective groups, the resulting free amino acid could be cyclized under high dilution conditions using diphenyl phosphorazidate (DPPA, (PhO)₂P(O)N₃)²⁴ and sodium hydrogen carbonate to produce (4*S*,4*R*)-antillatoxin (**1a**) in 45% yield (Scheme 5).

Although our synthetic (4S,5R)-antillatoxin showed reasonable ¹H and ¹³C NMR spectra and HRMS (EI, obsd M⁺ m/z503.3362, 0.5 ppm error for $C_{28}H_{45}N_3O_5$), its NMR spectra showed significant differences from those of the natural product. Furthermore, the optical rotation of the synthetic sample ($[\alpha]_D = -55$ (c 0.24, MeOH)) was also different from that of the natural product $([\alpha]_D = -140 \ (c \ 0.13,$ MeOH)). These differences led us to the conclusion that the formula **1a** does not accurately reflect the stereostructure for antillatoxin. On the basis of the assumption that the stereochemistries of amino acids are secure, we supposed that the stereochemistry at C4 and C5 would be misassigned. After the completion of our synthesis, White et al. also have accomplished the total synthesis of (4S,5R)antillatoxin (1a) and reached to the same conclusion as ours independently.^{11a} Unfortunately, the amount of natural





Scheme 7.

Scheme 6.

antillatoxin was very minute, and firm assignment of structure by a method other than synthesis might be difficult at this time.

Synthesis of (4*R*,5*R*)-antillatoxin (revised structure)

Next, we focused our attention on the synthesis of (4R,5R)antillatoxin, which was proposed as the second possible configuration by Gerwick et al.⁷ For the construction of the anti-C4, C5 configuration, we intended to use the anti-selective boron-mediated asymmetric aldol reaction developed by Abiko and Masamune.²⁵ After oxidation of alcohol **10**, the resulting aldehyde was added to the *E*-enolate solution, which was generated from the propionate ester of norephedrine derivative using dicyclohexylboron triflate and triethylamine. Subsequently, protection of the secondary alcohol gave the anti-aldol adduct **27**, which was converted to the corresponding alcohol **28** using DIBAL reduction (Scheme 6).

The absolute stereochemistry of chiral alcohol **28** was confirmed by comparison of the optical rotation data of the corresponding ketone **29** with **30**, which was derived from the already stereochemically evident alcohol **17** (Scheme 7).

The alcohol **28** was transformed into (4R,5R)-antillatoxin (**1b**) in the same way as developed in the synthesis of



 $[\alpha]_D$ -147 (c 0.23, MeOH) $[\alpha]_D$ -140 (c 0.13, MeOH) (natural product)



Figure 3. Revision of the proposed structure of antillatoxin.

(4*S*,5*R*)-antillatoxin (1a). Thus, the TPAP oxidation of 28 and Still's olefination gave the α , β -unsaturated ester 31, which was treated with TFA to afford the α , β -unsaturated lactone 32. Although the lactone 32 could be transformed into the phenylselenyl derivative 33, no stereoselectivity was observed in this 1,4-addition. Alkaline hydrolysis of 33, allyl esterification, and coupling with the tripeptide unit 2 produced the ester 34, which underwent the oxidation to give the linear precursor 35. Deprotection at the *N*- and *C*-terminals followed by macrolactamization with DPPA finally afforded (4*R*,5*R*)-antillatoxin (1b) (Scheme 8).

The synthetic (4R,5R)-antillatoxin (**1b**) was found to be identical to the natural antillatoxin by comparison of their ¹H NMR, ¹³C NMR and IR. The optical rotation of our synthetic sample ($[\alpha]_D = -147$ (*c* 0.23, MeOH)) also



Conclusion

In conclusion, we achieved the total synthesis of (4S,5R)and (4R,5R)-antillatoxins in an enantioselective convergent manner using Evans and Abiko–Masamune asymmetric aldol reactions, respectively. Our total synthesis indicates that the structure assigned to antillatoxin must be revised to the (4R,5R)-configuration (Fig. 3).

Furthermore, we have also accomplished the total synthesis of (4R,5S)- and (4S,5S)-antillatoxins by a common strategy which uses antipodal aldol reactions (Scheme 9 and Scheme





Scheme 10.

10). Biological and pharmacological evaluation of our synthetic four isomers of antillatoxin are currently underway.

Experimental

General information

Melting points were measured with a YANACO melting point apparatus (hot plate) and are uncorrected. Infrared spectra were recorded on a JASCO IRA-2 or SHIMADZU FT IR-8100 spectrometer. Optical rotations were measured on a JASCO DIP-140 or DIP-1000 digital polarimeters with a sodium lump (λ =589 nm, D line) and are reported as follows: [α]^T_D=(c g/100 ml, solvent).

¹H NMR spectra were recorded on a JEOL EX-270 (270 MHz) spectrometer. Chemical shifts are reported in ppm from tetramethylsilane as the internal standard. Data are reported as follows: chemical shift, integration, multiplicity (s=singlet, d=doublet, t=triplet, q=quartet, br= broad, m=multiplet), coupling constants (Hz), and assignment. Antillatoxin numbering is used for assignments on all intermediates. ¹³C NMR spectra were recorded on a JEOL EX-270 (67.8 MHz) spectrometer with complete proton decoupling. Chemical shifts are reported in ppm from tetramethylsilane with the solvent as the internal standard (deuterochloroform: δ 77.0 ppm).

Analytical thin layer chromatography was performed on Merck Art. 5715, Kieselgel $60F_{254}/0.25$ mm thickness

plates. Visualization was accomplished with UV light, phosphomolybdic acid, or ninhydrin solution followed by heating. Preparative thin layer chromatography was performed on Merck Art. 5744, Kiselgel 60F₂₅₄/0.5 mm thickness plates. Elementary analysis (Anal) and high-resolution mass spectra (HRMS) were done at the Analytical Facility at Nagoya City University.

Solvents for extraction and chromatography were reagent grade. Liquid chromatography was performed with forced flow (flash chromatography of the indicated solvent mixture on silica gel BW-820MH or BW-200 (Fuji Davison Co.)). Tetrahydrofuran (THF) was distilled from sodium metal/benzophenone ketyl. Dichloromethane (CH₂Cl₂), and hexamethylphosphoramide (HMPA) were distilled from calcium hydride. Acetonitrile (CH₃CN) and *N*,*N*-dimethyl-formamide (DMF) were dried over 4 Å molecular sieves. Triethylamine was dried over potassium hydroxide. All other commercially available reagents were used as received.

Boc-(*S*)-**Me-Val-Gly-OEt** (4). To a solution of Boc-(*S*)-Me-Val-OH (2.07 g, 8.96 mmol) and HCl·H-Gly-OEt (1.31 g, 9.39 mmol) in DMF (20 ml) was successively added dropwise DEPC (1.5 ml, 9.89 mmol) and triethylamine (3.1 ml, 22.2 mmol). After being stirred at 0°C for 2 h and then at room temperature for 3 h, the reaction mixture was diluted with ether and washed with 1 M aqueous KHSO₄, water, saturated aqueous NaHCO₃ and brine. The organic layer was dried (MgSO₄), filtered and concentrated. The residue was purified by silica gel column chromatography (BW-820MH, hexane–EtOAc=3:1) to

afford the desired product **4** as a colorless oil (2.444 g, 86%): $[\alpha]_D^{26} = -117.7$ (*c* 1.0, CHCl₃); IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹ 3337, 1755, 1682, 1679, 1526, 1480, 1445, 1393, 1368, 1196, 1157; ¹H NMR (CDCl₃) δ 0.87 (3H, d, *J*=6.9 Hz, Val-CH₃), 0.96 (3H, d, *J*=6.3 Hz, Val-CH₃), 1.27 (3H, t, *J*=6.9 Hz, CH₂CH₃), 2.28 (1H, m, Val-(CH₃)₂CH), 1.48 (9H, s, *t*-*Bu*), 2.80 (3H, s, N-CH₃), 4.00 (2H, d, *J*=5.6 Hz, Gly-CH₂), 4.19 (3H, m, Val- α -H, CH₂CH₃), 6.65 (1H, br, Gly-NH); ¹³C NMR (CDCl₃) δ 14.0, 18.4, 19.6, 26.0, 26.2, 28.3, 30.0, 41.0, 61.1, 64.1, 156.9, 169.4, 170.9. Anal. Calcd for C₁₅H₂₈N₂O₅: C, 56.94; H, 8.92; N, 8.85. Found: C, 56.75; H, 8.99; N, 8.71.

Alloc-(S)-Ala-(S)-Me-Val-Gly-OEt (5). The dipeptide 4 (503 mg, 1.59 mmol) was dissolved in CHCl₃ (2 ml), and TFA (2 ml) was added at room temperature. The solution was stirred for 20 min and concentrated. The residue was azeotropically concentrated with toluene $(\times 2)$. The resulting residue and Alloc-(S)-Ala-OH (275 mg, 1.59 mmol) were dissolved in CH₂Cl₂ (5.3 ml) and cooled to 0°C. BopCl 1.75 mmol) and triethylamine (445 mg, (0.67 ml. 4.8 mmol) were successively added, and the mixture was stirred at 0°C for 18 h. After dilution with EtOAc, the mixture was washed with 1 M aqueous KHSO₄, water, saturated aqueous NaHCO₃ and brine. The organic layer was dried (Na₂SO₄), filtered and concentrated. The residue was purified by silica gel column chromatography (BW-200, hexane-EtOAc=3:2) to afford the desired product 5 as a colorless oil (294 mg, 50%): $[\alpha]_D^{26} = -139.7$ (c 1.1, CHCl₃); IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹ 3316, 1723, 1684, 1638, 1530, 1464, 1244, 1200; ¹H NMR (CDCl₃) δ 0.84 (3H, d, J=6.6 Hz, Val-CH₃), 0.98 (3H, d, J=6.3 Hz, Val-CH₃) 1.26 (3H, t, J=7.3 Hz, CH₂CH₃), 1.35 (3H, d, J=6.6 Hz, Ala-CH₃), 2.33 (1H, m, Val-(CH₃)₂CH), 3.03 (3H, s, N-CH₃), 3.83 (1H, dd, J=17.8, 5.3 Hz, Gly-CH₂), 4.08 $(1H, dd, J=17.8, 6.6 Hz, Gly-CH_2), 4.18 (2H, q,$ J=7.3 Hz, CH₂CH₃), 4.53 (3H, m, CH₂=CHCH₂, Val-α-*H*), 4.68 (1H, m, Ala- α -*H*), 5.34-5.19 (2H, m, CH2=CHCH2), 5.66 (1H, d, J=7.6 Hz, Ala-NH), 5.98-5.80 (1H, m, $CH_2 = CHCH_2$), 6.55 (1H, br, Gly-NH); ¹³C NMR (CDCl₃) δ 14.0, 18.4, 19.4, 25.5, 30.4, 40.9, 47.1, 61.2, 62.4, 65.5, 117.5, 132.7, 155.4, 169.5, 170.1, 174.0. HRMS (EI) *m*/*z* Calcd for C₁₇H₂₉N₃O₆: 371.2056. Found: 371.2055.

Alloc-(S)-Ala-(S)-Me-Val-Gly-OH (2). To a solution of the tripeptide 5 (270 mg, 0.71 mmol) in THF (2 ml) at 0°C was added a solution of LiOH (60 mg) in water (2 ml). The resulting mixture was stirred at 0°C for 10 min. After dilution with water, the mixture was washed with ether. The aqueous layer was acidified to pH 3 by the addition of 1 M aqueous KHSO₄ and salted out. The mixture was extracted with ether $(\times 3)$. The combined organic extracts were dried (MgSO₄), filtered and concentrated to afford the desired product 2 as a colorless viscous oil (238 mg, 95%): $[\alpha]_{D}^{25} = -151.0 \ (c \ 1.0, \ CHCl_3); \ IR \ \nu_{max}^{neat} \ cm^{-1} \ 3316, \ 1720, \ 1600 \ max^{-1} \$ 1682, 1636, 1588, 1470, 1414, 1246, 1065; ¹H NMR $(CDCl_3) \delta 0.84 (3H, d, J=6.6 Hz, Val-CH_3), 0.97 (3H, d, d)$ J=6.6 Hz, Val-CH₃), 1.30 (3H, d, J=6.9 Hz, Ala-CH₃), 2.31 (1H, m, Val-(CH₃)₂CH), 3.03 (3H, s, N-CH₃), 3.94 (1H, m, Val- α -H), 4.11 (1H, m, Ala- α -H), 4.56 (2H, d, J=5.6 Hz, CH₂=CHCH₂), 4.65 (2H, d, J=11.2 Hz, Gly-CH₂), 5.23 (2H, m, CH₂=CHCH₂), 5.80–5.40 (1H, br, CO₂H), 5.85 (1H, m, CH₂=CHCH₂), 6.94 (1H, br, Gly-NH), 6.94 (1H, d, J=8.3 Hz, Ala-NH); ¹³C NMR (CDCl₃) δ 17.9, 18.5, 19.4, 25.7, 30.6, 40.9, 47.3, 62.8, 65.8, 117.8, 132.7, 155.7, 170.2, 172.3, 174.7. HRMS (EI) *m*/*z* Calcd for C₁₅H₂₅N₃O₆: 343.1743. Found: 343.1743.

Ethyl (2E)-2,4,4-trimethyl-2-pentenoate (8). To a solution of the phosphonate 7 (2.7 g, 10.1 mmol) in THF (30 ml) at -20° C was added *n*-BuLi (1.68 mol 1⁻¹ in hexane, 6 ml, 10.1 mmol). After being stirred at -20° C for 30 min, to the resulting solution was added pivaladehyde (1 ml, 9.21 mmol) by syringe. The reaction mixture was stirred at -20° C for 1 h and then at room temperature for 9.5 h. The reaction was quenched by the addition of saturated aqueous NH₄Cl and the mixture was extracted with brine, dried (MgSO₄), filtered and concentrated. The residue was distilled to afford 8 (922 mg, 59%) as a colorless oil, which was used for the next step without further purification: bp 120°C/25 mmHg (Kugelrohr); IR ν_{max}^{neat} cm⁻¹ 1711, 1644, 1466, 1366, 1281, 1250, 1200, 1109; ¹H NMR (CDCl₃) δ 1.18 (9H, s, t-Bu), 1.29 (3H, t, J=6.9 Hz, CH₂CH₃), 1.95 (3H, d, J=1.3 Hz, C_{15} -CH₃), 4.18 (2H, q, J=6.9 Hz, CH₂CH₃), 6.79 (1H, q, J=1.3 Hz, C_9 -H); ¹³C NMR (CDCl₃) δ 13.1, 14.1, 29.9, 32.8, 60.4, 126.5, 150.9, 169.1.

Ethyl (2E,4E)-2,4,6,6-tetramethyl-2,4-heptadienoate (9). To a solution of the ester 8 (705 mg, 4.14 mmol) in CH_2Cl_2 (10 ml) at $-78^{\circ}C$ was added DIBAL (0.95 mol/l in hexane, 13 ml, 12.35 mmol) dropwise. The resulting solution was stirred at $-78^{\circ}C$ for 50 min and quenched by the addition of 1 M aqueous KHSO₄. The mixture was extracted with ether (×1). The organic extracts were washed with 1 M aqueous KHSO₄, water, saturated aqueous NaHCO₃, water and brine. The organic layer was dried (MgSO₄), filtered and concentrated to afford the alcohol (631 mg) as a colorless oil, which was used for the next step without further purification.

To a solution of the alcohol (631 mg) in CH_2Cl_2 (11 ml) was added CMD (3 g, 34.5 mmol) in one portion. After being stirred at room temperature for 2.5 h, an additional CMD (3 g, 34.5 mmol) was added. After stirring for an additional 3.5 h, an additional CMD (2.5 g, 28.8 mmol) was added. The mixture was stirred for an additional 11 h, and CMD was removed by filtration through a Celite and washed with ether. The resulting filtrate was concentrated to afford the aldehyde (604 mg) as a colorless oil, which was used for the next step without further purification.

To a suspension of LiCl (218 mg, 5.14 mmol) in CH₃CN (6 ml) were subsequently added the phosphonate 7' (1.1 ml, 5.13 mmol) and DBU (0.7 ml, 4.68 mmol). After being stirred at room temperature for 5 min, to the resulting solution was added the above aldehyde in CH₃CN (2 ml, plus 2 ml rinse) by cannula. The mixture was stirred at room temperature for 8 h, and the bulk of solvent was removed. The residue was diluted with ether, washed with saturated NH₄Cl, water and brine. The organic layer was dried (MgSO₄), filtered and concentrated. Silica gel column chromatography (BW-820MH, hexane-ether=60:1) afforded the diene 9 as a colorless oil (330 mg, 38%). A small amount of the sample was purified for analysis by distillation (bp 130°C/10 mmHg, Kugelrohr): IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻

1767

1709, 1624, 1466, 1447, 1366, 1254, 1223, 1113; ¹H NMR (CDCl₃) δ 1.16 (9H, s, *t-Bu*), 1.30 (3H, t, *J*=6.9 Hz, CH₂CH₃), 1.91 (3H, d, *J*=1.3 Hz, C₁₄-CH₃) 1.97 (3H, d, *J*=1.3 Hz, C₁₅-CH₃), 4.20 (2H, q, *J*=6.9 Hz, CH₂CH₃), 5.56 (1H, br, C₉-H), 7.08 (1H, s, C₇-H); ¹³C NMR (CDCl₃) δ 13.8, 14.2, 17.2, 30.6, 32.9, 60.4, 125.1, 130.8, 144.8, 145.2, 169.1. Anal. Calcd for C₁₃H₂₂O₂: C, 74.24; H, 10.54. Found: C, 74.21; H, 10.68.

(2E,4E)-2,4,6,6-tetramethyl-2,4-heptadienol (10). By DIBAL reduction: To a solution of the ester 8 (281 mg, 4.14 mmol) in CH₂Cl₂ (10 ml) at -78° C was added DIBAL (1.5 mol 1⁻¹ in toluene, 2.7 ml, 4.05 mmol) dropwise. The resulting solution was stirred at -78° C for 25 min and quenched by the addition of 1 M aqueous KHSO₄. The mixture was extracted with CH₂Cl₂ (×3). The combined organic extracts were washed with saturated aqueous NaHCO₃, dried (Na₂SO₄), filtered and concentrated. The residue was purified by silica gel column chromatography (BW-820MH, hexane–ether=5:1) to afford the alcohol **10** as colorless oil (235 mg, quant.).

By Suzuki coupling: 4, 4-Dimethyl-2-pentyne (11) (1.44 g, 15 mmol) was dissolved in THF (20 ml) and catecholborane (1 M in THF, 15 ml, 15 mmol) was added. The mixture was heated to reflux for 1 h, washed with H₂O, and concentrated. The residue was diluted with THF (20 ml) and a solution of vinyl iodide 13 (1.39 g, 7 mmol) in THF (3 ml) was added. To this solution was added Cs_2CO_3 (4.56 g, 14 mmol), PdCl₂ (dppf)₂·CH₂Cl₂ (143 mg, 0.175 mmol) and AsPh₃ (107 mg, 0.35 mmol). After being stirred for 24 h at room temperature, the mixture was filtered through celite pad, and the filtrate was concentrated. The residue was purified by silica gel column chromatography (BW-820MH, hexaneether=5:1) to afford the desired product 10 as a colorless oil (1.178 g, quant.): IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹ 3339, 1466, 1445, 1362, 1071, 1009; H NMR (CDCl₃) δ 1.14 (9H, s, t-Bu), 1.77 $(3H, d, J=1.7 \text{ Hz}, C_{14}\text{-}CH_3), 1.82 (3H, d, J=1.3 \text{ Hz}, 1.82 \text{ (}3H, d, J=1.3 \text{ Hz}, 1.3 \text{ Hz}, 1.3$ C₁₅-CH₃), 2.53 (1H, br, OH), 3.99 (2H, br, CH₂), 5.31 (1H, s, C₉-H), 5.84 (1H, s, C₇-H); 13 C NMR (CDCl₃) δ 15.0, 17.8, 30.9, 32.4, 69.0, 130.8, 131.1, 133.7, 140.2. HRMS (EI) m/z Calcd for C₁₁H₂₀O: 168.1514. Found: 168.1487.

[4*R*,3(2*R*,3*R*)]-4-Benzyl-3-(3-hydroxy-2,4,6,8,8-pentamethyl-4,6-nonadienoyl)-2-oxazolidinone (15). To a solution of the alcohol 10 (644 mg, 3.83 mmol) in CH_2Cl_2 (11 ml) was added CMD (5.7 g, 65.6 mmol) in one portion. The mixture was stirred at room temperature for 4 h, and CMD was removed by filtration through a pad of celite and washed with ether. The resulting filtrate was concentrated to afford the aldehyde.

To a solution of propionyl oxazolidinone **14** (893 mg, 3.83 mmol) in CH₂Cl₂ (8 ml) at 0°C were added dropwise dibutylboryl triflate (1.14 ml, 4.53 mmol) and triethylamine (0.71 ml, 5.09 mmol) at a rate such that the internal temperature stayed below 3°C. The resulting clear, colorless solution was cooled to -78° C, and a solution of the above aldehyde in CH₂Cl₂ (1 ml, plus 1 ml rinse) was added by cannula. After 20 min, the solution was allowed to warm to -5° C for 40 min and quenched by the addition of the phosphate buffer (1.7 ml, pH 7) and methanol (4.8 ml). To this

was added 2:1 methanol-30% aqueous H₂O₂ (3.9 ml) carefully so as to keep the internal temperature below 5°C. The volatiles were removed in vacuo and water was added. The mixture was extracted with ether $(\times 1)$. The organic extracts were washed with saturated aqueous NaHCO₃ and brine, dried (MgSO₄), filtered and concentrated. The residue was purified by silica gel column chromatography (BW-200, hexane-EtOAc=6:1) to afford the desired product 15 as a yellow oil (1.429 g, 93%): $[\alpha]_{D}^{26} = -21.3$ (c 1.0, CHCl₃); IR ν_{max}^{neat} cm⁻¹ 3538, 1782, 1700, 1455, 1381, 1362, 1210, 1107; ¹H NMR (CDCl₃) δ 1.14 (9H, S, *t-Bu*), 1.21 (3H, d, J=6.9 Hz, C₁₃-CH₃), 1.72 (3H, d, J=1.3 Hz, C₁₄-CH₃), 1.81 (3H, d, J=1.3 Hz, C₁₅-CH₃), 2.80 (1H, dd, J=13.2, 9.6 Hz, ArCH₂), 2.81 (1H, d, J=3.0 Hz, OH), 3.28 (1H, dd, J=13.5, 3.3 Hz, ArCH₂), 4.01 (1H, m, C₁₃-H), 4.21 (2H, m, CH₂O), 4.37 (1H, br, C₅-H), 4.67 (1H, m, CHN), 5.28 (1H, s, C₉-H), 5.99 (1H, s, C₇-H), 7.34–7.20 (5H, m, Ar*H*); ¹³C NMR (CDCl₃) δ 10.6, 14.8, 18.0, 30.9, 32.5, 37.7, 40.5, 55.2, 66.1, 75.6, 127.3, 128.9, 129.3, 130.8, 131.6, 132.6, 135.0, 140.1, 152.9, 177.0. HRMS (EI) m/z Calcd for C₂₄H₃₃NO₄: 399.2409. Found: 399.2401.

Methyl (2R,3R,4E,6E)-3-hydroxy-2,4,6,8,8-pentamethyl-4,6-nonadienoate (16). To a solution of the aldol adduct 15 (1.429 g, 3.58 mmol) in THF-water (4:1, 8 ml) was added 30% aqueous H₂O₂ (1.45 ml, 14.3 mmol), followed by the addition of a solution of LiOH (240 mg, 5.72 mmol) in water (5.9 ml) at 0°C. After the solution was stirred for 25 min, sodium sulfite (1.8 g, 14.3 mmol) in water (10 ml) was added. The mixture was washed with ether, and the aqueous layer was acidified to pH 3 by the addition of 1 M aqueous KHSO₄. The mixture was salted out, and extracted with ether $(\times 3)$. The combined organic extracts were dried (MgSO₄), filtered and concentrated to afford the hydroxy acid. The hydroxy acid was dissolved in DMF (5 ml), and KHCO₃ (657 mg, 6.56 mmol) was added, followed by the addition of methyl iodide (0.37 ml, 5.94 mmol). The mixture was stirred at room temperature for 18 h. After dilution with ether, the mixture was washed with 1 M aqueous KHSO₄, water, saturated aqueous NaHCO₃ and brine. The organic layer was dried (MgSO₄), filtered and concentrated. The residue was purified by silica gel column chromatography (BW-820MH, hexane-EtOAc(5:1) to afford the desired product 16 as a colorless oil (542 mg, 60%): $[\alpha]_D^{26} = +8.7$ (c 1.1, CHCl₃); IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹ 3496, 1740, 1456, 1435, 1362, 1256, 1200, 1165, 1038; ¹H NMR (CDCl₃) δ 1.14 (9H, s, *t-Bu*), 1.16 (3H, d, *J*=7.3 Hz, C₁₃-CH₃), 1.71 (3H, d, J=1.3 Hz, C₁₄-CH₃), 1.79 (3H, d, J=7.3 Hz, C₁₅-CH₃), 2.59 (1H, br, OH), 2.75 (1H, m, C₄-H), 3.67 (3H, s, CH₃ ester), 4.25 (1H, br, C₅-H), 5.26 (1H, t, J=1.3 Hz, C_9 -H), 5.88 (1H, s, C_7 -H); ¹³C NMR (CDCl₃) δ 11.3, 13.8, 17.8, 30.9, 32.5, 43.1, 51.6, 76.5, 130.6, 132.2, 133.1, 140.1, 175.8. HRMS (EI) m/z Calcd for C₁₅H₂₆O₃: 254.1882. Found: 254.1882.

(2*S*,3*R*,4*E*,6*E*)-3-trimethylsiloxy-2,4,6,8,8-pentamethyl-4,6-nonadienol (17). To a solution of the alcohol 16 (524 mg, 2.13 mmol) in CH_2Cl_2 (7 ml) was added triethylamine (0.6 ml, 4.30 mmol), TESC1 (0.54 ml, 3.22 mmol) and DMAP (6 mg, 0.049 mmol) at 0°C. After being stirred at 0°C for 15 min and then at room temperature for 2.5 h, the mixture was diluted with ether. The mixture was washed with 1 M aqueous KHSO₄, water, saturated aqueous NaHCO₃, and brine. The organic layer was dried (MgSO₄), filtered and concentrated to afford the silyl ether.

To a solution of the silvl ether in CH₂Cl₂ was added DIBAL (0.94 M in hexane, 6.8 ml, 6.39 mmol) at -78°C . After being stirred at -78°C for 10 min and then at 0°C for 20 min, the reaction mixture was quenched by the addition of 1 M aqueous KHSO₄. The mixture was extracted with ether (×1). The organic extracts were washed with water, saturated aqueous NaHCO₃ and brine, dried (MgSO₄), filtered and concentrated. The residue was purified by silica gel column chromatography (BW-200, hexane-EtOAc= 6:1) to afford the desired product 17 as a colorless oil (712 mg, 98%): $[\alpha]_D^{25} = +1.9$ (*c* 1.1, MeOH); IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻ 3368, 1460, 1414, 1377, 1362, 1238, 1019; ¹H NMR (CDCl₃) δ 0.60 (6H, q, J=7.9 Hz, SiCH₂CH₃), 0.90 (3H, d, J=6.9 Hz, C₁₃-CH₃), 0.95 (9H, t, J=7.9 Hz, SiCH₂CH₃), 1.14 (9H, s, t-Bu), 1.71 (3H, d, J=1.3 Hz, C₁₄-CH₃), 1.79 (3H, d, J=1.3 Hz, C₁₅-CH₃), 1.88 (1H, m, C₄-H), 1.97 (1H, br, OH), 3.57-3.44 (2H, m, C₃-CH₂), 3.93 (1H, d, J=6.3 Hz, C_5 -*H*), 5.24 (1H, t, *J*=1.3 Hz, C_9 -*H*), 5.78 (1H, s, C_7 -*H*); ¹³C NMR (CDCl₃) δ 4.8, 6.8, 12.7, 13.9, 17.9, 31.0, 32.5, 39.8, 66.0, 80.8, 130.7, 132.1, 135.4, 139.8. Anal. Calcd for C₂₀H₄₀O₂Si: C, 70.52; H, 11.84. Found: C, 70.27; H, 12.12.

Methyl (4*S*,5*R*,2*Z*,6*E*,8*E*)-5-triethylsiloxy-4,6,8,10,10-pentamethyl-6,8-undecadienoate (18). To a stirred mixture of the alcohol 17 (525 mg, 1.54 mmol), *N*-methylmorpholine *N*-oxide (271 mg, 2.31 mmol) and powdered 4 Å molecular sieves (770 mg) in CH₃CN (6 ml) was added TPAP (27 mg, 0.077 mmol) in one portion at 0°C. After being stirred at 0°C for 5 min and then at room temperature for 20 min, the mixture was filtered through silica gel column and the filtrate was concentrated to afford the aldehyde.

A solution of bis(2,2,2-trifluoroethyl)(methoxycarbonylmethyl)phosphonate (0.65 ml, 3.07 mmol) and 18-crown-6 (1.93 g, 7.30 mmol) in THF (10 ml) was cooled to -78° C and treated with KHMDS (0.5 M in toluene, 5.8 ml, 2.9 mmol). A solution of the aldehyde in THF (2 ml, plus 2 ml rinse) was added by cannula and the resulting mixture was stirred at -78°C for 20 min and then at -10°C for 20 min. The reaction mixture was quenched by the addition of saturated aqueous NH₄Cl and the mixture was extracted with ether $(\times 1)$. The organic extracts were washed with brine, dried (MgSO₄), filtered and concentrated. The residue was purified by silica gel column chromatography (BW-200, hexane-ether=50:1) to afford the desired product **18** as a colorless oil (449 mg, 74%): $[\alpha]_{D}^{26} = +86.0$ $(c 1.2, \text{CHCl}_3)$; IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹ 1727, 1646, 1460, 1437, 1408, 1379, 1362, 1237, 1196, 1177, 1073, 1009; ¹H NMR (CDCl₃) δ 0.57 (6H, q, J=8.3 Hz, SiCH₂CH₃), 0.94 (9H, t, J=8.3 Hz, SiCH₂CH₃), 1.01 (3H, d, J=6.6 Hz, C₁₃-CH₃), 1.13 (9H, s, t-Bu), 1.64 (3H, d, J=1.3 Hz, C₁₄-CH₃), 1.75 (3H, d, *J*=1.0 Hz, C₁₅-CH₃), 3.67 (1H, m, C₄-H), 3.71 (3H, s, CH₃ ester), 3.79 (1H, d, 6.9 Hz, C₅-H), 5.18 (1H, t, J=1.3 Hz, C₉-H), 5.68 (1H, dd, J=11.6, 0.7 Hz, C₂-H), 5.73 (1H, s, C₇-*H*), 6.02 (1H, dd, *J*=11.5, 10.6 Hz, C₃-*H*); ¹³C NMR (CDCl₃) δ 4.9, 6.9, 13.2, 15.7, 17.8, 31.0, 32.5, 37.3, 51.0, 81.8, 117.7, 130.8, 131.7, 135.8, 139.6, 153.3, 166.7. Anal. Calcd for C₂₃H₄₂O₃Si: C, 70.00; H, 10.73. Found: C, 69.93; H, 10.67.

(4S,5R)-5-((1E,3E)-1,3,6,6-Tetramethyl-1,3-hexadienyl)-4-methyl-2-pentene-5-olide (19). To a solution of the ester 18 (449 mg, 1.14 mmol) in CH_2Cl_2 (7 ml) was added dropwise TFA (0.053 ml, 0.69 mmol) at -10° C. The mixture was slowly warmed to room temperature over 19 h, and then the additional TFA (0.015 ml, 0.20 mmol) was added. After stirring at room temperature for 3.5 h, the reaction mixture was quenched by the addition of triethylamine. After concentration of the mixture, the residue was purified by silica gel column chromatography (BW-200, hexane-EtOAc=5:1) to afford the desired product **19** as a colorless oil (187 mg, 66%): $[\alpha]_D^{25} = +357.6$ (c 0.3, CHCl₃); IR ν_{max}^{neat} cm⁻¹ 1728, 1453, 1381, 1372, 1246, 1107, 1065; ¹H NMR (CDCl₃) δ 0.96 (3H, d, J=7.3 Hz, C₁₃-CH₃), 1.15 (9H, s, *t-Bu*), 1.74 (3H, s, C_{14} -CH₃), 1.82 (3H, d, J=1.0 Hz, C₁₅-CH₃), 2.56 (1H, m, C₄-H), 4.80 (1H, br, C₅-H), 5.30 $(1H, t, J=1.3 \text{ Hz}, C_9-H), 6.00 (1H, d, J=9.6, C_2-H), 6.12$ (1H, s, C₇-*H*), 7.02 (1H, dd, J=9.6 Hz, 6.3 Hz, C₃-*H*); ¹³C NMR (CDCl₃) δ 11.6, 14.8, 17.7, 30.7, 31.6, 32.4, 82.1, 119.6, 127.9, 130.3, 131.9, 140.2, 151.7, 164.1. HRMS (EI) *m*/*z* Calcd for C₁₆H₂₄O₂: 248.1776. Found: 248.1765.

(3R,4S,5R)-5-((1E,3E)-1,3,6,6-Tetramethyl-1,3-hexadienyl)-4-methyl-3-(phenylseleno)methyl-5-pentenolide (20). To a solution of $(PhSe)_2CH_2$ (192 mg, 0.589 mmol) in THF (1.1 ml) was slowly added *n*-BuLi (1.61 M in hexane, 0.39 ml, 0.628 mmol) at -78° C, and the resulting solution was stirred at the same temperature for 1.5 h. After addition of HMPA (1 ml), a solution of the lactone 19 (112 mg, 0.453 mmol) in THF (0.2 ml, plus 0.1 ml rinse) was added by cannula. The mixture was stirred at -78° C for 30 min, and then quenched by the addition of saturated aqueous NH_4Cl . The mixture was extracted with ether (×1), and the organic extracts were washed with brine, dried (MgSO₄), filtered and concentrated. The residue was purified by silica gel column chromatography (BW-200, hexane-EtOAc=6:1) to afford the desired product 20 as white crystals (138 mg, 73%): mp 80–82°C (ether– pentane); $[\alpha]_D^{24}$ =+32.8 (*c* 0.6, CHCl₃); IR $\nu_{\text{max}}^{\text{KBr disk}}$ cm⁻¹ 1728, 1578, 1478, 1439, 1375, 1256, 1080; ¹H NMR (CDCl₃) δ 0.84 (3H, d, J=6.9 Hz, C₁₃-CH₃), 1.14 (9H, s, *t-Bu*), 1.65 (3H, s, C_{14} -CH₃), 1.79 (3H, d, J=1.0 Hz, C₁₅-CH₃), 2.10 (1H, m, C₃-H), 1.94 (1H, m, C₄-H), 2.41 (1H, dd, J=16.2, 9.6 Hz, C_2 -H), 2.68 (1H, dd, J=16.2, 6.6 Hz, C_2 -H), 3.03 (2H, d, J=6.6 Hz, PhSeCH₂), 4.65 (1H, br, C_5 -H), 5.26 (1H, t, J=1.3 Hz, C_9 -H), 6.00 (1H, s, C₇-H), 7.27 (3H, m, ArH), 7.52 (2H, m, ArH); ¹³C NMR (CDCl₃) δ 14.3, 15.2, 17.9, 30.9, 32.5, 34.0, 34.3, 35.4, 37.9, 81.2, 127.4, 127.9, 129.3, 130.5, 132.0, 132.9, 133.0, 140.3, 171.7. HRMS (EI) *m/z* Calcd for C₂₃H₃₂O₂Se: 420.1567. Found: 420.1568.

Vinyl iodide (22). To a solution of the alcohol **21** (206 mg, 0.725 mmol) in CH₂Cl₂ (2.5 ml) was added triethylamine (0.2 ml, 1.43 mmol), TESC1 (0.18 ml, 1.07 mmol) and DMAP (5 mg, 0.041 mmol) at 0°C. After being stirred at 0°C for 5 min and then at room temperature for 1.5 h, the mixture was diluted with ether. The reaction mixture was washed with 1 M aqueous KHSO₄, water, saturated aqueous NaHCO₃ and brine. The organic layer was dried (MgSO₄), filtered and concentrated to afford the silyl ether.

To a solution of the silvl ether in CH₂Cl₂ (2.5 ml) was added

DIBAL (0.94 M in hexane, 2.3 ml, 2.16 mmol) at -78° C. After being stirred at -78° C for 10 min and then at -10° C for 10 min, the reaction mixture was quenched by the addition of 1 M aqueous KHSO₄. The mixture was extracted with ether $(\times 1)$. The organic extracts were washed with water, saturated aqueous NaHCO₃, and brine, dried, (MgSO₄), filtered and concentrated. The residue was purified by silica gel column chromatography (BW-200, hexane-EtOAc=6:1) to afford the desired product 22 as a colorless oil (254 mg, 88%): $[\alpha]_D^{24} = +33.6$ (*c* 1.1, CHCl₃); IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹ 3368, 1456, 1414, 1379, 1264, 1240; ¹H NMR (CDCl₃) δ 0.56 (6H, q, J=8.3 Hz, SiCH₂CH₃), 0.84 (3H, d, J=6.6 Hz, C₁₃-CH₃), 0.92 (9H, t, J=8.3 Hz, SiCH₂CH₃), 1.77 (3H, s, C₁₄-CH₃), 1.88 (1H, m, C₄-H), 2.01 (1H, br, OH), 3.39–3.55 (2H, m, C₃-CH₂), 4.21 (1H, d, J=5.3 Hz, C₅-*H*), 6.17 (1H, d, J=1.0 Hz, C₇-*H*); ¹³C NMR (CDCl₃) δ 4.6, 6.5, 11.6, 21.0, 39.4, 65.2, 78.1, 78.4, 149.0. Anal. Calcd for C₁₃H₂₇IO₂Si: C, 42.16; H, 7.35. Found: C, 42.20; H, 7.56.

(2S,3R,4E,6E)-3-trimethylsiloxy-2,4,6,8,8-pentamethyl-4,6-nonadienol (17) (by Suzuki coupling). 4,4-Dimethyl-2-pentyne (11) (0.060 ml, 0.45 mmol) was dissolved in THF (0.9 ml) and catecholborane (1 M in THF, 0.45 ml, 0.45 mmol) was added. The mixture was heated to reflux for 1.5 h and concentrated. The residue was diluted with THF (0.8 ml) and a solution of vinyl iodide 22 (83 mg, 0.208 mmol) in THF (0.2 ml, plus 0.1 ml rinse) was added by cannula. To this solution was added 2 N aqueous NaOH (0.46 ml) and Pd(Ph₃P)₄ (12 mg, 0.010 mmol). The reaction mixture was heated to reflux for 2 h, and quenched by the addition of 4 N aqueous NaOH (0.1 ml) and 30% aqueous H_2O_2 (0.1 ml). After being stirred at room temperature for 1 h, the mixture was extracted with ether $(\times 3)$. The combined organic extracts were washed with brine, dried (MgSO₄), filtered, and concentrated. The residue was purified by silica gel column chromatography (BW-820MH, hexane-ether=6:1) to afford the desired product 17 as a yellow oil (43 mg, 61%). ¹H and ¹³C NMR spectra were identical with those of the authentic sample.

Ester (23). To a solution of the lactone 20 (70 mg, 0.167 mmol) in THF (1.1 ml) was added a solution of LiOH (21 mg, 0.500 mmol) in water at 0°C. The resulting mixture was stirred at 0°C for 1.5 h. After dilution with ether, the mixture was washed with 1 M aqueous KHSO₄ and brine. The organic layer was dried (MgSO₄), filtered, and concentrated to afford the hydroxy acid.

The hydroxy acid was dissolved in DMF (1.1 ml), and KHCO₃ (51 mg, 0.51 mmol) was added, followed by the addition of allyl bromide (0.029 mmol, 0.335 mmol). The mixture was stirred at room temperature for 3 h. After dilution with ether, the mixture was washed with 1 M aqueous KHSO₄, water, saturated aqueous NaHCO₃, and brine. The organic layer was dried (MgSO₄), filtered and concentrated to afford the allyl ester.

The allyl ester and tripeptide unit 2 (124 mg, 0.352 mmol) were dissolved in CH_2Cl_2 (1.4 mol) and EDCI-HCl (74 mg, 0.386 mmol) was added, followed by the addition of DMAP (2 mg, 0.016 mmol) at 0°C. After being stirred at 0°C for 15 min and then at room temperature for 13 h, the mixture

was diluted with ether. The mixture was washed with 1 M aqueous KHSO₄, water, saturated aqueous NaHCO₃ and brine. The organic layer was dried (MgSO₄), filtered, and concentrated. The residue was purified by silica gel column chromatography (BW-200, hexane-EtOAc=2:1~1:1) to afford the desired product 23 as a colorless oil (104 mg, 78%): $[\alpha]_{D}^{24} = -79.8$ (c 0.7, CHCl₃); IR ν_{max}^{neat} cm⁻¹ 3324, 1732, 1686, 1638, 1478, 1414, 1377, 1246, 1188; ¹H NMR (CDCl₃) δ 0.84 (3H, d, J=6.6 Hz, Val-CH₃), 0.85 $(3H, d, J=6.9 \text{ Hz}, C_{13}-CH_3), 0.97 (3H, d, J=6.6 \text{ Hz},$ Val-CH₃), 1.14 (9H, s, t-Bu), 1.33 (3H, d, J=6.9 Hz, Ala-CH₃), 1.66 (3H, d, J=1.0 Hz, C₁₄-CH₃), 1.74 (3H, d, J=1.0 Hz, C₁₅-CH₃), 2.08 (1H, m, C₄-H), 2.25 (1H, m, C3-H), 2.32 (1H, m, Val-(CH3)2CH), 2.37 (1H, m, C2-H), 2.67 (1H, dd, *J*=11.9, 10.6 Hz, C₂-*H*), 2.81 (1H, dd, *J*=15.8, 3.3, PhSeCH₂), 3.01 (3H, s, N-CH₃), 3.05 (1H, dd, J=11.9, 3.3 Hz, PhSeCH₂), 3.79 (1H, dd, J=18.2, 5.0 Hz, Gly-CH₂), 3.95 (1H, m, Val- α -H), 4.07 (1H, dd, J=18.1, 6.6 Hz, Gly-CH₂), 4.55 (4H, d, J=5.6 Hz, CH₂=CHCH₂×2), 4.65 $(1H, m, Ala - \alpha - H)$, 5.09 $(1H, d, J = 8.9 \text{ Hz}, C_5 - H)$, 5.26 $(4H, d, J = 8.9 \text{ Hz}, C_5 - H)$ m, CH₂=CHCH₂×2), 5.33 (1H, s, C₉-H), 5.63 (1H, br, Ala-NH), 5.72 (1H, s, C₇-H), 5.97–5.83 (2H, m, CH2=CHCH2×2), 6.50 (1H, br, Gly-NH), 7.24 (3H, m, ArH), 7.47 (2H, m, ArH); ¹³C NMR (CDCl₃) δ 10.2, 12.9, 17.8, 18.2, 18.4, 19.5, 25.3, 29.0, 30.4, 30.8, 32.6, 36.5, 36.6, 36.8, 41.0, 47.1, 62.6, 65.1, 65.6, 83.3, 117.6, 118.3, 127.1, 129.0, 129.8, 130.0, 130.2, 132.0, 132.7, 136.1, 141.1, 156.7, 168.6, 170.0, 172.1, 174.0. Anal. Calcd for C₄₁H₆₁N₃O₈Se: C, 61.33; H, 7.66; N, 5.23. Found: C, 61.41; H, 7.79; N, 51.4.

Protected linear peptide (25). To a solution of the ester 23 (36 mg, 0.044 mmol) in THF (0.3 ml)-water (0.2 ml) was added NaIO₄ (28 mg, 0.131 mmol). The mixture was stirred at room temperature for 41.5 h, and directly purified by silica gel chromatography (BW-200, hexane-EtOAc=2:1) to afford the desired product 25 as a colorless oil (27 mg 93%): $[\alpha]_{\rm D}^{24} = -77.3$ (c 0.7, CHCl₃); IR $\nu_{\rm max}^{\rm neat}$ cm⁻¹ 3320, 1732, 1692, 1640, 1464, 1246, 1190; ¹H NMR (CDCl₃) δ 0.84 (3H, d, J=6.6 Hz, Val-CH₃), 0.97 (3H, d, J=6.6 Hz, Val-CH₃), 1.05 (3H, d, J=6.9 Hz, C₁₃-CH₃), 1.12 (9H, s, *t-Bu*), 1.34 (3H, d, *J*=6.9 Hz, Ala-CH₃), 1.68 (3H, d, J=1.3 Hz, C₁₄-CH₃), 1.75 (3H, d, J=1.3 Hz, C₁₅-CH₃), 2.25 (1H, m, C₃-H), 2.32 (1H, m, C₄-H), 2.65 (1H, q, J=6.9 Hz, Val-(CH₃)₂CH), 3.02 (3H, s, N-CH₃), 3.05 (2H, s, C₂-CH₂), 3.85 (1H, dd, J=18.1, 5.3 Hz, Gly-CH₂), 4.09 $(1H, dd, J=18.2, 6.3 Hz, Gly-CH_2), 4.57 (4H, m,$ CH2=CHCH2×2), 4.65 (2H, m, Ala-α-H, Val-α-H), 5.03 (2H, s, C₁₂-*H*), 5.35–5.14 (6H, m, CH₂=CHCH₂×2, C₅-*H*, C₉-H), 5.66 (1H, br, Ala-NH), 5.76 (1H, s, C₇-H), 5.98-5.79 (2H, m, CH₂=CHCH₂×2), 6.55 (1H, br, Gly-NH); ¹³C NMR (CDCl₃) δ 14.0, 15.1, 17.8, 18.3, 18.4, 19.6, 25.4, 30.5, 30.9, 32.6, 40.4, 40.9, 41.5, 47.2, 62.7, 65.4, 65.6, 81.9, 115.8, 117.7, 118.4, 130.2, 130.3, 132.0, 132.7, 134.5, 140.8, 142.9, 155.4, 168.6, 170.0, 171.0, 174.0. HRMS (EI) m/z Calcd for C₃₅H₅₅N₃O₈: 645.3989. Found: 645.3989.

(45,5*R*)-Antillatoxin (1a). To a solution of the ester 25 (26 mg, 0.040 mmol) in THF (0.4 ml) was added morpholine (0.070 ml, 0.803 mmol) and $Pd(Ph_3P)_4$ (4.6 mg, 0.004 mmol). After being stirred at room temperature for 40 min, the mixture was diluted with the phosphate buffer

(pH 6) and extracted with $CHCl_3$ (×3). The organic extracts were dried (Na₂SO₄), filtered, and concentrated to afford the amino acid.

The amino acid was dissolved in DMF (20 ml) and cooled to 0°C. NaHCO₃ (24 mg, 0.286 mmol) and DPPA (0.018 ml, 0.083 mmol) were added, and the solution was stirred at 0°C for 3 days. After concentration below 40°C, the residue was diluted with EtOAc and the organic phase was washed with 1 M aqueous KHSO₄, water, saturated aqueous NaHCO₃, and brine. The organic layer was dried (Na₂SO₄), filtered and concentrated. Preparative thin layer chromatography (0.5 mm thickness, CHCl₃-MeOH=20:1) followed by silica gel column chromatography (BW-200, hexane-EtOAc=1:1) afforded the desired product 1a as a colorless oil (9 mg, 0.018 mmol, 45%): $[\alpha]_D^{25} = -55.2$ (c 0.24, MeOH); IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹ 3287, 1740, 1688, 1644, 1624, 1549, 1462, 1275; ¹H NMR (CDCl₃) δ 0.84 (3H, d, J=6.9 Hz, Val-CH₃), 0.97 (3H, d, J=7.6 Hz, C₁₃-CH₃), 0.98 (3H, d, J=6.3 Hz, Val-CH₃), 1.12 (9H, s, t-Bu), 1.38 (3H, d, J=6.6 Hz, Ala-CH₃), 1.64 (3H, d, J=2.0 Hz, C₁₄-CH₃), 1.74 (3H, d, J=1.3 Hz, C₁₅-CH₃), 2.41 (1H, m, Val-(CH₃)₂CH), 2.72 (1H, m, C₄-H), 2.90 (1H, d, J=7.3 Hz, C₂-H), 2.92 (3H, s, N-CH₃) 3.48 (1H, d, J=7.3 Hz, C₂-H), 3.58 (1H, dd, J=17.5, 1.7 Hz, Gly-CH₂), 4.16 (1H, d, J=10.6 Hz, Val-α-H), 4.71 (1H, dd, J=17.5, 9.6 Hz, Gly-CH₂), 5.20 (1H, s, C₉-H), 5.21 (3H, m, C₅-H, C₁₂-H), 5.38 (1H, m, Ala-α-H), 5.55 (1H, s, C₇-H), 6.41 (1H, d, J=9.9 Hz, Ala-NH), 7.99 (1H, d, J=8.9 Hz, Gly-NH); ¹³C NMR (CDCl₃) δ 14.7, 15.7, 18.0, 18.7, 19.4, 26.9, 28.8, 30.9, 32.6, 40.7, 41.6, 42.6, 44.5, 67.0, 80.5, 119.7, 129.8, 130.4, 131.8, 140.4, 144.6, 168.0, 168.2, 171.3, 171.7. HRMS (EI) *m*/*z* Calcd for C₂₈H₄₅N₃O₅: 503.3359. Found: 503.3362.

(1*S*,*2R*)-2-(*N*-Benzyl-*N*-mesitylenesulfonyl)amino-1-phenyl-1-propyl(2*S*,*3S*,4*E*,6*E*)-3-triethylsiloxy-2,4,6,8,8-pentamethyl-4,6-nonadienoate (27). To a solution of the alcohol 10 (1.35 g, 8.0 mmol) in CH_2Cl_2 (24 ml) was added CMD (11.7 g, 136 mmol) in one portion. The mixture was stirred at room temperature for 4 h, and CMD was removed by filtration through a pad of Celite and washed with Et_2O . The resulting filtrate was concentrated to afford the aldehyde, which was used for the next step without further purification.

To a stirred solution of 26 (4.03 g, 8.4 mmol) in CH₂Cl₂ (80 ml) was added Et₃N (2.69 ml, 19.2 mmol). The solution was cooled to -78° C and to this was added via cannula a solution of c-Hex₂BOTf (0.9 M in hexane, 18.7 ml, 16.8 mmol) in CH₂Cl₂ (2 ml), which was precooled to -78° C. This resulting solution was stirred at -78° C for 2 h to complete enolization. The aldehyde was added dropwise to the solution, and the reaction mixture was stirred at -78° C for 1 h and at 0°C for 1 h. The reaction mixture was quenched by the addition of the phosphate buffer (32 ml, pH 7), followed by MeOH (80 ml) and 30% H_2O_2 (8 ml). The mixture was stirred overnight vigorously, the volatiles were removed in vacuo, and water was added. The mixture was extracted with Et_2O (×1) and the organic extracts were washed with saturated aqueous NaHCO₃ and brine, dried (MgSO₄), filtered, and concentrated to afford the aldol adduct, which was used for the next step without further purification. The aldol adduct was dissolved in CHCl₃ (16 ml), and 2,6lutidine (3.48 ml, 30.0 mmol) was added, followed by the addition of TESOTf (54.3 ml, 24 mmol). The mixture was stirred at 0°C for 4 h. After dilution with Et₂O, the mixture was washed with 1 M aqueous KHSO₄ and brine. The organic layer was dried (MgSO₄), filtered and concentrated. The residue was purified by silica gel column chromatography (BW-820 MH, hexane-EtOAc=4:1) to afford the desired product 27 as a colorless oil (5.47 g, 90%): $[\alpha]_{D}^{25} = -56.2$ (c 1.0, CHCl₃); IR ν_{max}^{neat} cm⁻¹ 1748, 1682, 1605, 1497, 1456, 1416, 1379, 1329, 1258, 1235, 1206, 1156, 1013, 930, 907, 884, 858, 808, 749, 730; ¹H NMR (CDCl₃) δ 0.53 (6H, q, *J*=7.9 Hz, SiCH₂CH₃), 0.81 (3H, d, J=7.3 Hz, C₁₃-CH₃), 0.90 (9H, t, J=7.9 Hz, SiCH₂CH₃), 1.10 (3H, d, J=6.9 Hz, N CHCH₃), 1.14 (9H, s, t-Bu), 1.71 (3H, s, C₁₄-CH₃), 1.77 (3H, d, J=0.99 Hz, C₁₅-CH₃), 2.33 (3H, s, ArCH₃), 2.47 (6H, s, C₁₅-CH₃), 2.73 (1H, m, C_4 -H), 3.96 (1H, m, NCH), 4.11 (1H, d, J=9.6 Hz, C_5 -H), 4.42 (1H, A of AB, J=16.2 Hz, PhCH₂), 4.98 (1H, B of AB, J=16.2 Hz, PhCH₂), 5.24 (1H, s, C₉-H), 5.68 (1H, d, J=4.6 Hz, PhCH), 5.74 (1H, s, C₇-H), 6.65 (2H, d, J=6.9 Hz, ArH), 6.93 (2H, s, ArH), 7.11 (4H, m, ArH), 7.25 (2H, m, ArH), 7.47 (2H, m, ArH); ¹³C NMR (CDCl₃) δ 4.8, 6.9, 11.7, 14.4, 14.4, 17.7, 22.8, 30.1, 30.9, 32.6, 44.9, 48.4, 56.8, 77.2, 81.2, 126.0, 127.2, 127.7, 128.2, 128.3, 128.4, 130.5, 132.1, 133.2, 133.3, 134.8, 138.4, 139.0, 140.3, 140.4, 142.4, 174.0. Anal. Calcd for C₄₅H₆₅NO₅SSi: C, 71.10; H, 8.62; N 1.84. Found: C, 70.91; H, 8.80; N 1.64.

(2R,3R,4E,6E)-3-Trimethylsiloxy-2,4,6,8,8-pentamethyl-4,6-nonadienol (28). To a solution of the silvl ether 27 (3.66 g, 4.8 mmol) in CH₂Cl₂ (100 ml) was added DIBAL (1.5 M in toluene, 9.7 ml, 14.5 mmol) at -78° C. After being stirred at -78° C for 30 min, the reaction mixture was quenched by the addition of 1 M aqueous KHSO₄. The mixture was extracted with $Et_2O(\times 1)$. The organic extracts were washed with water, saturated aqueous NaHCO₃ and brine, dried (MgSO₄), filtered and concentrated. The residue was purified by silica gel column chromatography (BW-820) MH, hexane-EtOAc=5:1) to afford the desired product 28 as a colorless oil (1.64 g, quant.): $[\alpha]_{D}^{25} = -8.3$ (c 0.85, MeOH); IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹ 3432, 1462, 1416, 1379, 1362, 1237, 1202, 1057, 1007, 909, 889, 857, 808, 739, 727; ¹H NMR (CDCl₃) δ 0.62 (6H, q, J=7.6 Hz, SiCH₂CH₃), 0.75 $(3H, d, J=6.9 \text{ Hz}, C_{13}\text{-}CH_3), 0.96 (9H, t, J=7.6 \text{ Hz},$ SiCH₂CH₃), 1.14 (9H, s, t-Bu), 1.68 (3H, d, J=0.99 Hz, C₁₄-CH₃), 1.79 (3H, d, J=0.99 Hz, C₁₅-CH₃), 1.90 (1H, m, C₄-H), 3.23 (1H, m, OH), 3.61 (2H, m, C₃-CH₂), 3.83 (1H, d, J=8.3 Hz, C₅-H), 5.25 (1H, s, C₉-H), 5.74 (1H, s, C₇-H); ¹³C NMR (CDCl₃ δ 4.8, 6.8, 12.4, 14.2, 17.8, 31.0, 32.6, 38.3, 67.8, 58.9, 130.5, 133.3, 134.8, 140.1. Anal. Calcd for C₂₀H₄₀O₂Si: C, 70.52; H, 11.84. Found: C, 70.32; H, 11.69.

(2*R*,4*E*,6*E*)-1-Triethylsiloxy-3-oxo-2,4,6,8,8-pentamethyl-4,6-nonadienol (29). A mixture of 28 (130 mg, 0.38 mmol) and TBAF (1.0 M in THF, 0.95 ml, 0.95 mmol) in THF (1 ml) was stirred at 0°C for 1 h. After dilution with Et₂O, the mixture was washed with H₂O and saturated brine, dried (MgSO₄) and concentrated in vacuo. The residue was purified by silica gel column chromatography (BW-820 MH, hexane–EtOAc=5:1) to afford the diol as a colorless oil (80 mg, 93%): $[\alpha]_D^{23}$ -62.9 (*c* 0.5, CHCl₃); IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹ 3368, 1464, 1379, 1362, 1256, 1233, 1202, 1088, 1011, 982, 891, 847, 810; ¹H NMR (CDCl₃) δ 0.75 (3H, d, *J*=6.9 Hz, C₁₃-CH₃), 1,14 (9H, s, *t-Bu*), 1.75 (3H, d, *J*=0.99 Hz, C₁₄-CH₃), 1.82 (3H, d, *J*=1.32 Hz, C₁₅-CH₃), 1.95 (1H, m, C₄-H), 2.37 (1H, br, OH), 3.01 (1H, br, OH), 3.70 (2H, m, C₃-CH₂), 3.88 (1H, br, *J*=8.9 Hz, C₅-H), 5.31 (1H, s, C₉-H), 5.81 (1H, s, C₇-H); ¹³C NMR (CDCl₃) δ 12.3, 13.8, 17.9, 30.9, 32.5, 37.5, 68.4, 85.4, 130.5, 134.0, 134.9, 140.7. HRMS (EI) m/z Calcd for C₁₆H₂₆O₂: 226.1933. Found: 226.1950.

To a solution of the alcohol (50 mg, 0.22 mmol) in $CHCl_3$ (0.6 ml) was added triethylamine (0.034 ml, 0.24 mmol), TESC1 (0.040 ml, 0.24 mmol) and DMAP (2.4 mg, 0.020 mmol) at 0°C. After being stirred at 0°C for 15 min and then at room temperature overnight, the mixture was diluted with Et_2O . The mixture was washed with 1 M aqueous KHSO₄ and brine. The organic layer was dried (MgSO₄), filtered and concentrated to afford the silyl ether.

To a stirred mixture of the silvl ether, N-methylmorpholine *N*-oxide (54 mg, 0.0.46 mmol) and powdered 4 Å molecular sieves (154 mg) in CH₃CN (1.2 ml) was added TPAP (6 mg, 0.017 mmol) in one portion at 0°C. After being stirred at 0°C for 30 min and then at room temperature for 30 min, the mixture was filtered through silica gel and the filtrate was concentrated in vacuo. The residue was purified by silica gel column chromatography (BW-820 MH, hexane-EtOAc= 20:1) to afford 29 as a colorless oil (61 mg, 82%): $[\alpha]_{\rm D}^{23}$ = 23.0 (c 0.9, MeOH); IR $\nu_{\rm max}^{\rm neat}$ cm⁻¹ 1663, 1615, 1452, 1416, 1383, 1364, 1237, 1213, 1100, 1038, 1017, 810, 745; ¹H NMR (CDCl₃) δ 0.56 (6H, q, J=7.9 Hz, SiCH₂CH₃), 0.92 (9H, t, J=7.9 Hz, SiCH₂CH₃), 1.06 (3H, d, J=6.6 Hz, C₁₃-CH₃), 1,18 (9H, s, t-Bu), 1.91 (3H, d, J=1.3 Hz, C₁₄-CH₃), 1.94 (3H, d, J=1.3 Hz, C₁₅-CH₃), 3.52 (2H, m, C₃-H), 3.81 (1H, m, C₄-H), 5.59 (1H, s, C₉-*H*), 6.98 (1H, s, C₇-*H*); ¹³C NMR (CDCl₃) δ 4.3, 6.4, 14.9, 17.4, 30.7, 33.1, 42.2, 66.1, 77.2, 131.1, 134.5, 145.5, 145.7, 205.9. Anal. Calcd for C₂₀H₃₈O₂Si: C, 70.94; H, 11.31. Found: C, 70.66; H, 11.27.

(2*S*,4*E*,6*E*)-1-Triethylsiloxy-3-oxo-2,4,6,8,8-pentamethyl-4,6-nonadienol (30). The alcohol 17 (108 mg, 0.317 mmol) was treated as described for the synthesis of **29** to afford the diol as a colorless oil (107 mg, quant.): $[\alpha]_D^{23} = +18.3$ (*c* 1.4, CHCl₃); IR ν_{max}^{neat} cm⁻¹ 3368, 1464, 1379, 1362, 1254, 1233, 1202, 1178, 1116, 1088, 1036, 1013, 982, 895; ¹H NMR (CDCl₃) δ 0.95 (3H, d, *J*=7.3 Hz, C₁₃-CH₃), 1,15 (9H, s, *t*-*Bu*), 1.72 (3H, d, *J*=1.3 Hz, C₁₄-CH₃), 1.82 (3H, d, *J*=1.32 Hz, C₁₅-CH₃), 1.93 (1H, m, C₄-*H*, OH×2), 3.66 (2H, d, *J*=5.0 Hz, C₃-CH₂), 4.12 (1H, d, *J*=5.0 Hz, C₅-*H*), 5.29 (1H, s, C₉-*H*), 5.88 (1H, s, C₇-*H*); ¹³C NMR (CDCl₃) δ 10.8, 14.6, 18.1, 31.0, 32.6, 37.9, 66.8, 77.2, 130.8, 131.1, 135.2, 140.3. HRMS (EI) *m*/*z* Calcd for C₁₆H₂₆O₂: 226.1933. Found: 226.1945.

The ester (10 mg, 0.044 mmol) was treated as described for the synthesis of **29** to afford the ketone **30** as a colorless oil (10 mg, 67%): IR, ¹H NMR and ¹³C NMR spectra were identical with **29**. $[\alpha]_D^{23} = +21.5$ (*c* 0.5, MeOH). Anal. Calcd for C₂₀H₃₈O₂Si: C, 70.94; H, 11.31. Found: C, 70.63; H, 11.05.

Methyl (4R,5R,2Z,6E,8E)-5-Triethylsiloxy-4,6,8,10,10pentamethyl-6,8-undecadienoate (31). The alcohol 27 (1.02 g, 3.00 mmol) was treated as described for the synthesis of **18** to give the ester **31** as a colorless oil (966 mg, 82%): $[\alpha]_D^{21}$ =-35.8 (*c* 1.0, CHCl₃); IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹ 1727, 1654, 1458, 1437, 1408, 1377, 1362, 1223, 1194, 1175, 1130, 1102, 1075, 1034, 1007, 972, 858, 822, 741, 725; ¹H NMR (CDCl₃) δ 0.56 (6H, q, *J*=7.9 Hz, SiCH₂CH₃), 0.92 (12H, m, SiCH₂CH₃), 1.76 (3H, s, C₁₅-CH₃), 3.69 (3H, s, CH₃ ester), 3.76 (2H, m, C₄-H, C₅-H), 5.20 (1H, m, C₉-H), 5.73 (1H, s, C₂-H), 5.77 (1H, s, C₇-H), 6.17 (1H, dd, *J*=11.6, 9.6 Hz, C₃-H); ¹³C NMR (CDCl₃) δ 4.9, 6.9, 13.3, 17.4, 17.8, 31.0, 32.5, 37.4, 50.9, 82.4, 118.7, 130.8, 132.1, 135.3, 139.6, 153.6, 166.8. Anal. Calcd for C₂₃H₄₂O₂Si: C, 70.00; H, 10.70. Found: C, 69.70; H, 10.79.

(4*R*,5*R*)-5-((1*E*,3*E*)-1,3,6,6-Tetramethyl-1-3-hexadienyl)-4-methyl-2-pentene-5-olide (32). The ester 31 (600 mg, 1.5 mmol) was treated as described for the synthesis of 19 to afford the lactone 32 as a colorless oil (234 mg, 63%): $[\alpha]_{D}^{23} = -96.4$ (*c* 0.5, CHCl₃); IR ν_{max}^{neat} cm⁻¹ 1730, 1651, 1375, 1289, 1264, 1235, 1175, 1201, 1084, 1009, 918, 891, 847, 814, 733; ¹H NMR (CDCl₃) δ 1.02 (3H, d, *J*=7.3 Hz, C₁₃-CH₃), 1.13 (9H, s, *t*-*Bu*), 1.78 (3H, s, C₁₄-CH₃), 1.80 (3H, s, C₁₅-CH₃), 2.72 (1H, m, C₄-*H*), 4.33 (1H, d, *J*=11.2 Hz, C₅-*H*), 5.32 (1H, s, C₉-*H*), 5.88 (1H, s, C₂-*H*), 5.97 (1H, dd, *J*=5.9.9, 2.6 Hz, C₇-*H*), 6.68 (1H, dd, *J*=9.9, 2.6 Hz, C₃-*H*); ¹³C NMR (CDCl₃) δ 12.7, 15.7, 17.6, 30.8, 31.1, 32.6, 90.7, 120.1, 129.1, 130.1, 137.5, 141.4, 151.9, 164.5. HRMS (EI) *m*/*z* Calcd for C₁₆H₂₄O₂: 248.1776. Found: 248.1764.

Ester (34). The lactone 32 (154 mg, 0.623 mmol) was treated as described for the synthesis of 20 and 23 to afford the ester 34 as a colorless oil (77 mg, 15% in 4 steps): $[\alpha]_{D}^{23} = -35.4 \ (c \ 1.0, \ CHCl_{3}); \ IR \ \nu_{max}^{neat} \ cm^{-1} \ 3314, \ 1732, 1682, \ 1645, \ 1580, \ 1530, \ 1480, \ 1414, \ 1377, \ 1275, \ 1244,$ 1192, 1148, 1094, 1065, 1022, 994, 930, 750, 739; ¹H NMR (CDCl₃) δ 0.67 (3H, d, J=7.0 Hz, C₁₃-CH₃), 0.83 (3H, d, J=6.4 Hz, Val-CH₃), 0.97 (3H, d, J=6.4 Hz, Val-CH₃), 1.12 (9H, s, t-Bu), 1.33 (3H, d, J=7.0 Hz, Ala-CH₃), 1.66 (3H, d, J=1.22 Hz, C₁₄-CH₃), 1.76 (3H, d, J=1.22, C₁₅-CH₃), 2.18 (1H, m, C₄-H), 2.29–2.40 (2H, m, C₃-H, Val-(CH₃)CH), 2.46 (1H, m, C₂-CH₂), 2.78 (1H, m, C₂-CH₂), 2.86 (1H, s, PhSeCH₂), 3.02 (3H, s, N-CH₃), 3.05 (1H, dd, J=12.5, 5.8 Hz, PhSeCH₂), 3.76 (2H, dd, J=18.1, 5.3 Hz, Gly-CH₂), 3.86 (1H, dd, J=18.3, 5.5 Hz, Val-α-H), 4.57 (4H, m, CH₂=CHCH₂×2), 4.67 (1H, m, Ala-α-H), 4.91 (1H, d, J=9.5 Hz, C₅-H), 5.20-5.30 (4H, m, CH₂=CHCH₂×2), 5.32 (1H, d, J=1.5 Hz, C₉-H), 5.66 (1H, br, J=7.9 Hz, Ala-NH), 5.81 (1H, s, C₇-H), 5.84–5.95 (2H, m, CH₂=CHCH₂×2), 6.42 (1H, t, J=5.5 Hz, Gly-NH), 7.27 (3H, m, ArH), 7.51 (2H, m, ArH); ¹³C NMR (CDCl₃) δ 10.5, 13.0, 18.4, 18.5, 19.7, 25.6, 30.3, 30.5, 30.9, 32.6, 34.5, 34.7, 35.8, 40.9, 47.2, 62.7, 65.3, 65.6, 82.8, 117.6, 118.4, 127.1, 129.1, 129.9, 130.4, 132.2, 132.7, 133.0, 136.2, 141.2, 155.4, 168.7, 169.8, 172.4, 173.8. Anal. Calcd for C₄₁H₆₁N₃O₈Se·0.5EtOAc: C, 60.98; H, 7.74; N, 4.96. Found: C, 61.26; H, 7.80; N, 4.60.

Protected linear peptide (35). The ester **34** (16 mg, 0.02 mmol) was treated as described for the synthesis of **25** to give the protected linear peptide **35** as a colorless oil

(10 mg, 81%): $[\alpha]_{\rm D}^{23} = -43.6$ (c 0.95, CHCl₃); IR $\nu_{\rm max}^{\rm neat}$ cm⁻¹ 3346, 1735, 1692, 1640, 1534, 1466, 1414, 1375, 1364, 1327, 1273, 1244, 1194, 1152, 1084, 1063, 1036, 1013, 994, 930, 851, 820, 777, 756; ¹H NMR (CDCl₃) δ 0.83 $(3H, d, J=6.7 \text{ Hz}, \text{Val-C}H_3), 0.94 (3H, d, J=7.0),$ C₁₃-CH₃), 0.97 (3H, d, J=6.7, Val-CH₃), 1.13 (9H, s, t-Bu), 1.33 (3H, d, J=7.0 Hz, Ala-CH₃), 1.69 (3H, d, J=1.5 Hz, C₁₄-CH₃), 1.79 (3H, d, J=1.2 Hz, C₁₅-CH₃), 2.31 (1H, m, C₄-H), 2.61 (1H, m, Val-(CH₃)CH), 3.02 (3H, s, N-CH₃), 3.07 (2H, d, J=5.2 Hz, C₂-CH₂), 3.83 (1H, dd, J=18.1, 5.3 Hz, Gly-CH₂), 4.00 (1H, dd, J=18.3, 5.3 Hz, Gly-CH₂), 4.59 (6H, m, Val-α-H, CH₂=CHCH₂×2, Ala-α-H), 5.04 (3H, m, C₁₂-H, C₅-H), 5.20-5.35 (5H, m, C₉-H, CH₂=CHCH₂×2), 5.64 (1H, d, J=8.2 Hz, Ala-NH), 5.90 (1H, s, C₇-H), 5.92 (2H, m, CH₂=CHCH₂×2), 6.48 (1H, brt, J=5.48 Hz, Gly-NH); ¹³C NMR (CDCl₃) δ 12.8, 16.7, 17.8, 18.4, 18.5, 19.6, 25.5, 30.5, 30.9, 32.6, 40.6, 40.9, 41.7, 47.2, 62.7, 65.4, 65.6, 84.3, 115.3, 117.6, 118.5, 129.6, 130.3, 130.4, 132.7, 136.8, 141.3, 143.5, 155.4, 168.3, 169.8, 171.2, 173.9. HRMS (EI) m/z Calcd for C₃₅H₅₅N₃O₈: 645.3989. Found: 645.3990.

(4R,5R)-Antillatoxin (1b). The protected linear peptide 35 (18 mg, 0.027 mmol) was treated as described for the synthesis of 1a to afford the desired product 1b as a colorless oil (5 mg, 37%): $[\alpha]_D^{21} = -147$ (c 0.23, MeOH); IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹ 3281, 1738, 1692, 1646, 1553, 1260; ¹H NMR (CDCl₃) δ 0.86 (3H, d, J=6.9 Hz, Val-CH₃), 0.88 (3H, d, J=6.7 Hz, C₁₃-CH₃), 0.98 (3H, d, J=6.3 Hz, Val-CH₃), 1.12 (9H, s, t-Bu), 1.42 (3H, d, J=6.6 Hz, Ala-CH₃), 1.56 (3H, d, J=1.0 Hz, C₁₄-CH₃), 1.79 (3H, d, J=1.0 Hz, C₁₅-CH₃), 2.17 (1H, m, C₄-H), 2.45 (1H, m, Val-(CH₃)₂CH), 2.79 (1H, d, J=12.5 Hz, C₂-H), 2.87 (3H, s, N-CH₃), 2.97 (1H, d, J=13.2 Hz, C₂-H), 3.48 (1H, dd, J=18.2, 1.3 Hz, Gly- CH_2), 4.25 (1H, d, J=10.8 Hz, Val- α -H), 4.69 (1H, dd, J=18.2, 9.9 Hz, Gly-CH₂), 5.01 (1H, s, C₁₂-H), 5.06 (1H, s, C₁₂-H), 5.17 (1H, d, J=10.9 Hz, C₅-H), 5.30 (1H, s, C_9 -H), 5.36 (1H, m, Ala- α -H), 5.94 (1H, s, C_7 -H), 6.66 (1H, d, J=9.4 Hz, Ala-NH), 7.97 (1H, d, J=9.6 Hz, Gly-NH), 13 C NMR (CDCl₃) δ 12.4, 17.8, 18.6, 18.7, 18.9, 19.3, 26.1, 28.8, 30.9, 32.6, 39.0, 41.1, 43.1, 67.1, 83.4, 113.8, 129.2, 130.4, 137.3, 141.5, 144.8, 167.6, 167.8, 171.1, 173.2. HRMS (EI) *m/z* Calcd for C₂₈H₄₅N₃O₅: 503.3359. Found: 503.3362.

[4S,3(2S,3S)]-4-Benzyl-3-(3-hydroxy-2,4,6,8,8-pentamethyl-4,6-nonadienoyl)-2-oxazolidinone (36). The alcohol 10 (2.26 g, 13.4 mmol) was treated as described for the synthesis of 15 to afford 36 as a pale yellow oil (5.609 g, 98%): IR, ¹H NMR and ¹³C NMR were identical with 15. $[\alpha]_D^{26}$ =+19.7 (*c* 1.0, CHCl₃); Anal. Calcd for C₂₄H₃₃O₄·0.25H₂O: C, 71.35; H, 8.36; N 3.47. Found: C, 71.52; H, 8.26; N 3.14.

Methyl (2*S*,3*S*,4*E*,6*E*)-3-Hydroxy-2,4,6,8,8-pentamethyl-4,6-nonadienoate (37). To a solution of the aldol adduct 36 (1.80 g, 4.19 mmol) in CH₂Cl₂ (40 ml) was added NaOMe (0.4 M in MeOH, 11.5 ml, 4.6 mmol) at -30° C. After being stirred for 10 min, the mixture was quenched with 1 M aqueous KHSO₄, and extracted with Et₂O (×2). The combined organic extracts were dried (MgSO₄), filtered and concentrated. The residue was purified by silica gel column chromatography (BW-820 MH, hexane–EtOAc= 6:1) to afford **37** as a colorless oil (942 mg, 88%): $[\alpha]_D^{26} = -9.1$ (*c* 1.1, CHCl₃); IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹ 3475, 1740, 1658, 1458, 1437, 1377, 1362, 1348, 1256, 1200, 1165, 1121, 1169, 1040, 1017, 889, 857, 758, 712; ¹H NMR (CDCl₃) δ 1.14 (9H, s, *t-Bu*), 1.16 (3H, d, *J*=7.3 Hz, C₁₃-CH₃), 1.70 (3H, s, C₁₄-CH₃), 1.79 (3H, s, C₁₅-CH₃), 2.44 (1H, br, OH), 2.75 (1H, m, C₄-H), 3.68 (3H, s, CH₃ ester), 4.27 (1H, br, C₅-H), 5.26 (1H, s, C₉-H), 5.78 (1H, s, C₇-H); ¹³C NMR (CDCl₃) δ 11.3, 13.9, 18.0, 31.0, 32.6, 43.1, 51.8, 76.6, 130.7, 131.8, 132.3, 133.1, 140.2, 176.0. HRMS (EI) *m/z* Calcd for C₁₅H₂₆O₃: 254.1882. Found: 254.1879.

(2*R*,3*S*,4*E*,6*E*)-3-Trimethylsiloxy-2,4,6,8,8-pentamethyl-4,6-nonadienol (38). To a solution of the alcohol 37 (850 mg, 3.34 mmol) was treated as described for the synthesis of 17 to afford the alcohol 38 as a colorless oil (1.04 g, quant.): IR, ¹H NMR and ¹³C NMR spectra were identical with 17. $[\alpha]_D^{24} = -1.6$ (*c* 1.1, MeOH); Anal. Calcd for C₂₀H₄₀O₂Si: C, 70.52; H, 11.84. Found: C, 70.21; H, 11.75.

Methyl (4*R*,5*S*,2*Z*,6*E*,8*E*)-5-Triethylsiloxy-4,6,8,10,10-pentamethyl-6,8-undecadienoate (39). The alcohol 38 (1.08 mg, 3.17 mmol) was treated as described for the synthesis of 18 to afford the ester 39 as a colorless oil (907 mg, 73%): IR, ¹H NMR and ¹³C NMR spectra were identical with 18. $[\alpha]_D^{25}$ =-89.3 (*c* 0.85, CHCl₃); Anal. Calcd for C₂₃H₄₂O₃Si: C, 70.00; H, 10.73. Found: C, 69.78; H, 10.55.

(4*R*,5*S*)-5-((1*E*,3*E*)-1,3,6,6-Tetramethyl-1,3-hexadienyl)-4-methyl-2-pentene-5-olide (40). The ester 39 (237 mg, 0.70 mmol) was treated as described for the synthesis of 19 to afford the lactone 40 as a colorless oil (113 mg, 76%): IR, ¹H NMR and ¹³C NMR were identical with 19. $[\alpha]_D^{32}$ =-352.6 (*c* 0.3, CHCl₃); HRMS (EI) *m*/*z* Calcd for C₁₆H₂₄O₂: 248.1776. Found: 248.1775.

(3*R*,4*S*,5*R*)-5-((1*E*,3*E*)-1,3,6,6-Tetramethyl-1,3-hexadienyl)-4-methyl-3-(phenylseleno)methyl-5-pentenolide (41). The lactone 40 (96 mg, 0.389 mmol) was treated as described for the synthesis of 20 to afford the selenolactone 41 as a yellow oil (110 mg, 68%): IR, ¹H NMR and ¹³C NMR spectra were identical with 20. $[\alpha]_D^{24} = -31.1^\circ$ (*c* 0.6, CHCl₃); HRMS (EI) *m*/*z* Calcd for C₂₃H₃₂O₂Se: 420.1567. Found: 420.1568.

Ester (42). The lactone **41** (77 mg, 0.184 mmol) was treated as described for the synthesis of **23** to afford the ester **42** as a colorless oil (86 mg, 58%): $[\alpha]_D^{24} = -61.1$ (*c* 0.25, CHCl₃); IR ν_{max}^{neat} cm⁻¹ 3304, 1732, 1682, 1638, 1537, 1477, 1414, 1379, 1244, 1190, 1148, 1065, 1022, 990, 932, 891, 739; ¹H NMR (CDCl₃) δ 0.84 (3H, d, *J*=6.6 Hz, Val-CH₃), 0.85 (3H, d, *J*=6.9 Hz, C₁₃-CH₃), 0.97 (3H, d, *J*=6.6 Hz, Val-CH₃), 1.14 (9H, s, *t-Bu*), 1.32 (3H, d, *J*=6.6 Hz, Ala-CH₃), 1.66 (3H, s, C₁₄-CH₃), 1.74 (3H, s, C₁₅-CH₃), 2.07 (1H, m, C₄-H), 2.31–2.41 (3H, m, C₃-H, Val-(CH₃)₂CH, C₂-H), 2.67 (1H, m, C₂-H), 2.77 (1H, d, *J*=4.3 Hz, PhSeCH₂) 3.01 (3H, s, N-CH₃), 3.07 (1H, m, PhSeCH₂), 3.96 (3H, m, Gly-CH₂, Val-α-H), 4.55 (4H, m, CH₂=CHCH₂×2), 4.62 (1H, s, Ala-α-H), 5.09 (1H, d, *J*=8.6 Hz, C₅-H), 5.72 (2H, m, Ala-NH,

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C₇-*H*), 5.85–5.95 (2H, m, CH₂==C*H*CH₂×2), 6.57 (1H, br, Gly-N*H*), 7.24 (3H, m, Ar*H*), 7.47 (2H, m, Ar*H*); ¹³C NMR (CDCl₃) δ 10.2, 13.0, 17.8, 18.3, 18.4, 19.6, 25.5, 29.2, 30.5, 30.9, 32.6, 36.6, 36.7, 36.8, 41.0, 47.2, 62.6, 65.2, 65.6, 83.3, 117.6, 118.4, 127.2, 129.1, 129.8, 130.0, 130.3, 132.0, 132.7, 136.0, 141.1, 155.4, 168.7, 170.0, 172.1, 173.9. Anal. Calcd for C₄₁H₆₁N₃O₈Se: C, 61.33; H, 7.66; N, 5.23. Found: C, 61.44; H, 7.70; N, 4.95.

Protected linear peptide (43). The ester 42 (21 mg, 0.026 mmol) was treated as described for the synthesis of 25 to afford the protected linear peptide 43 as a colorless oil (16 mg, quant.): $[\alpha]_D^{25} = -82.0$ (c 0.54, CHCl₃); IR ν_{max}^{neat} cm⁻¹ 3324, 1738, 1732, 1684, 1636, 1507, 1456, 1375, 1364, 1326, 1273, 1244, 1192, 1150, 1090, 1063, 936, 932, 777, ¹H NMR (CDCl₃) δ 0.84 (3H, d, J=6.6 Hz, Val-CH₃), 0.98 (3H, d, J=6.6 Hz, Val-CH₃), 1.04 (3H, d, J=6.9 Hz, C_{13} -CH₃), 1.12 (9H, s, t-Bu), 1.33 (3H, d, J=6.9 Hz, Ala-CH₃), 1.68 (3H, s, C₁₄-CH₃), 1.75 (3H, s, C₁₅-CH₃), 2.32 (1H, m, C₄-H), 2.65 (1H, m, Val-(CH₃)₂CH), 3.00 (3H, s, N-CH₃), 3.05 (2H, s, C₂-CH₂) 3.97 (1H, dd, J=18.2, 5.0 Hz, Gly-CH₂), 4.07 (1H, dd, J=178, 5.9 Hz, Gly-CH₂), 4.56 (4H, m, CH₂=CHCH₂×2), 4.61 (2H, m, Ala-α-H, Val-α-H), 5.02 (2H, s, C₁₂-H), 5.16-5.34 (6H, m, CH2=CHCH2×2, C5-H, C9-H), 5.62 (1H, br, Ala-N*H*), 5.77 (1H, s, C₇-*H*), 5.83–5.98 (2H, m, CH_2 =C*H*CH₂×2), 6.45 (1H, br, Gly-N*H*); ¹³C NMR (CDCl₃) & 14.1, 14.8, 17.8, 18.3, 18.4, 19.6, 25.4, 30.5, 30.9, 32.6, 40.5, 40.9, 41.4, 47.2, 62.8, 65.4, 65.6, 81.7, 115.8, 117.7, 118.4, 130.1, 130.4, 132.0, 132.7, 134.3, 140.7, 143.0, 155.3, 168.6, 169.9, 171.0, 173.9. MS *m*/*z*: 645 (M⁺).

(4R,5S)-Antillatoxin (1c). The protected linear peptide 43 (23 mg, 0.0356 mmol) was treated as described for the synthesis of 1a to afford the desired product 1c as a colorless oil (6 mg, 37%): $[\alpha]_D^{24} = -64.8$ (c 0.13, MeOH); IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹ 3291, 1744, 1686, 1626, 1534, 1464, 1262; ¹H NMR $(DMSO-d_6) \delta 0.76 (3H, d, J=6.9 Hz, Val-CH_3), 0.90 (3H, d, d)$ J=7.3 Hz, C_{13} -CH₃), 1.00 (3H, d, J=6.3 Hz, Val-CH₃), 1.11 (9H, s, *t-Bu*), 1.18 (3H, d, *J*=6.3 Hz, Ala-CH₃), 1.63 (3H, s, C₁₄-CH₃), 1.73 (3H, d, J=1.0 Hz, C₁₅-CH₃), 2.18–2.24 (2H, m, C₄-H, Val-(CH₃)₂CH), 2.64 (3H, s, N-CH₃), 2.94 (1H, s, C_2 -H), 3.04 (1H, s, C_2 -H), 3.67 (1H, dd, J=16.8, 5.0 Hz, Gly-CH₂), 3.93 (1H, d, J=9.6 Hz, Val-α-H), 4.18 (1H, dd, J=16.8, 6.9 Hz, Gly-CH₂), 4.64 (1H, m, Ala- α -H), 4.78 (1H, s, C₅-H), 4.94, (1H, s, C₁₂-H), 5.00 (1H, s, C₁₂-H), 5.20 (1H, s, C₉-H), 5.61 (1H, s, C₇-H), 8.00 (1H, br, Gly-NH), 8.35 (1H, d, J=8.3 Hz, Ala-NH); ¹³C NMR (DMSO-d₆) δ 14.0, 15.6, 17.7, 17.9, 18.7, 27.1, 29.0, 30.8, 32.2, 40.9, 41.7, 42.2, 44.9, 64.9, 81.2, 124.9, 130.4, 130.6, 139.6, 144.7, 167.9, 169.1, 169.4, 171.4. HRMS (EI) m/z Calcd for C₂₈H₄₅N₃O₅:503.3359. Found: 503.3362.

[1*R*, 2*S*)]-2-(*N*-Benzyl-*n*-mesitylenesulfonyl)amino-1-phenyl-1-propyl(2*R*, 3*R*, 4*E*, 6*E*)-3-triethylsiloxy-2,4,6,8,8-pentamethyl-4,6-nonadienoate (44). The alcohol 10 (13.5 g, 8.0 mmol) was treated as described for the synthesis of 27 to afford the aldol adduct 44 as a colorless oil (5.59 g, 92%): IR ¹H NMR and ¹³C NMR spectra were identical with 27. $[\alpha]_D^{21}$ =+54.1 (*c* 1.0, CHCl₃); Anal. Calcd for C₄₅H₆₅NO₅SSi: C, 71.10; H, 8.62; N 1.84. Found: C, 70.98; H, 8.65; N 1.59. (2*S*,3*S*,4*E*,6*E*)-3-Trimethylsiloxy-2,4,6,8,8-pentamethyl-4,6-nonadioenol (45). The aldol adduct 44 (4.03 g, 5.3 mmol) was treated as described for the synthesis of 28 to afford the alcohol 45 as a pale yellow oil (1.81 g, quant.): IR ¹H NMR and ¹³C NMR spectra were identical with 28. $[\alpha]_D^{25}=+8.8$ (*c* 1.0, MeOH). Anal. Calcd. for C₂₀H₄₀NO₂Si: C, 70.52; H, 11.84. Found: C, 70.26; H, 11.75.

Methyl (4*S*,5*S*,2*Z*,6*E*,8*E*)-5-Triethylsiloxy-4,6,8,10,10-pentamethyl-6,8-undecadienoate (46). The alcohol 45 (788 mg, 2.30 mmol) was treated as described for the synthesis of 31 to afford the ester 46 as a colorless oil (724 mg, 80%): IR, ¹H NMR and ¹³C NMR spectra were identical with 31. $[\alpha]_D^{21}$ =+34.0 (*c* 1.0, CHCl₃); Anal. Calcd for C₂₃H₄₂O₂Si·0.5H₂O: C, 68.43; H, 10.74. Found: C, 68.69; H, 10.47.

(4*S*,5*S*)-5-((1*E*,3*E*)-1,3,6,6-Tetramethyl-1,3-hexadienyl)-4-methyl-2-pentene-5-olide (47). The ester 46 (600 mg, 1.5 mmol) was treated as described for the synthesis of 32 to afford the lactone 47 as a colorless oil (200 mg, 54%): IR, ¹H NMR and ¹³C NMR spectra were identical with 32. $[\alpha]_D^{23}$ =+96.1 (*c* 0.4, CHCl₃). HRMS (EI) *m/z* Calcd for C₁₆H₂₄O₂: 248.1776. Found: 248.1783.

Ester (49). The lactone 47 (50 mg, 0.20 mmol) was treated as described for the synthesis of 34 to afford the ester 49 as a colorless oil (30 mg, 18%): $[\alpha]_D^{23} = -43.6$ (*c* 0.33, CHCl₃); IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹ 3325, 3090, 1732, 1684, 1651, 1522, 1478, 1456, 1418, 1377, 1275, 1244, 1190, 1148, 1065, 1022, 995, 930, 779, 739; ¹H NMR (CDCl₃) δ 0.66 (3H, d, J=6.9 Hz, C₁₃-CH₃), 0.82 (3H, d, J=6.6 Hz, Val-CH₃), 0.96 (3H, d, J=6.3 Hz, Val-CH₃), 1.12 (9H, s, t-Bu), 1.30 (3H, d, J=6.1 Hz, Ala-CH₃), 1.66 (3H, s, C₁₄-CH₃), 1.75 (3H, s, C₁₅-CH₃, 2.16 (1H, m, C4-H), 2.28–2.46 (3H, m, C3-H, Val-(CH₃)CH, C₂-CH₂), 2.75-2.84 (2H, m, C₂-H, PhSeCH₂), 2.98 (3H, s, N-CH₃), 3.02 (1H, m, PhSeCH₂), 3.50 (2H, dd, J=14.0, 4.1 Hz, Gly-CH₂), 4.01 (1H, dd, J=18.1, 6.9 Hz, Val- α -H), 4.53 (4H, m, CH₂=CHCH₂×2), 4.62 (1H, m, Ala- α -H), 4.88 (1H, d, J=10.2 Hz, C₅-H), 5.19-5.34 (5H, m, CH2=CHCH2×2, C9-H), 5.63 (1H, d, J=7.9 Hz, Ala-NH), 5.81 (1H, s, C₇-H), 5.83-5.96 (2H, m, CH₂=CHCH₂×2), 6.42 (1H, t, J=5.3 Hz, Gly-NH), 7.27 (3H, m, ArH), 7.51 (2H, m, ArH); ¹³C NMR (CDCl₃) δ 10.3, 12.9, 17.8, 18.4, 19.6, 25.3, 30.3, 30.9, 31.5, 32.6, 34.3, 34.5, 35.8, 40.7, 47.2, 62.5, 65.4, 65.6, 82.7, 117.6, 118.4, 127.1, 129.7, 129.9, 130.3, 132.1, 132.7, 133.0, 136.4, 141.1, 155.4, 168.7, 169.8, 172.5, 173.8. Anal. Calcd for C41H61N3O8Se 0.5EtOAc: C, 60.98; H, 7.74; N, 4.96. Found: C, 61.26; H, 7.80; N, 4.16.

Protected linear peptide (50). The ester **49** (20 mg, 0.025 mmol) was treated as described for the synthesis of **35** to afford the protected linear peptide **50** as a colorless oil (12 mg, 80%): $[\alpha]_D^{23} = -52.4$ (*c* 1.26 CHCl₃); IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹ 3325, 3090, 1735, 1686, 1640, 1526, 1466, 1414, 1380, 1367, 1333, 1282, 1242, 1192, 1152, 1063, 994, 930; ¹H NMR (CDCl₃) δ 0.82 (3H, d, *J*=6.6 Hz, Val-CH₃), 0.95 (6H, m, C₁₃-CH₃, Val-CH₃), 1.13 (9H, s, *t*-Bu), 1.32 (3H, d, *J*=6.9 Hz, Ala-CH₃), 1.68 (3H, d, *J*=0.99 Hz, C₁₄-CH₃), 1.78 (3H, d, *J*=0.99 Hz, C₁₅-CH₃), 2.30 (1H, m, C₄-H), 2.59 (1H, m, Val-(CH₃)CH), 3.00 (3H, s, N-CH₃), 3.05 (2H, m,

C₂-CH₂), 3.82 (1H, dd, J=18.2, 5.3 Hz, Gly-CH₂), 4.00 (1H, dd, J=180, 6.1 Hz, Gly-CH₂), 4.58 (6H, m, Val- α -H, CH₂=CHCH₂×2, Ala- α -H), 5.03 (3H, m, C₁₂-H, C₅-H), 5.18–5.36 (5H, m, C₉-H, CH₂=CHCH₂×2), 5.64 (1H, d, J=8.3 Hz, Ala-NH), 5.90 (3H, m, C₇-H, CH₂=CH₂×2), 6.51 (1H, m, Gly-NH); ¹³C NMR (CDCl₃) δ 12.7, 16.7, 17.8, 19.6, 25.5, 30.4, 30.9, 32.6, 40.9, 41.8, 47.2, 62.6, 65.4, 65.6, 84.1, 115.3, 117.6, 118.5, 129.5, 130.4, 132.1, 136.8, 141.2, 143.6, 155.4, 168.3, 169.8, 171.2, 173.9. MS m/z: 645 (M⁺).

(4S,5S)-Antillatoxin (1d). The protected linear peptide 50 (8 mg, 0.012 mmol) was treated as described for the synthesis of 1b to afford the desired product 1d as a colorless oil (2 mg, 32%): $[\alpha]_D^{25} = -8.7$ (c 0.1, MeOH); IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹ 3304, 1744, 1682, 1622, 1547, 1538, 1471, 1464, 1456, 1446, 1412, 1375, 1361, 1335, 1271, 1175, 1121, 1075, 1013, 974, 906, 754; ¹H NMR (CDCl₃) δ 0.85 (3H, d, J=7.0 Hz, Val-CH₃), 0.90 (3H, d, J=7.0 Hz, C_{13} -CH₃), 0.99 (3H, d, J=6.7 Hz, C₄-H), 1.14 (9H, s, t-Bu), 1.38 (3H, d, J=6.7 Hz, Ala-CH₃), 1.78 (3H, s, C₁₄-CH₃), 1.88 (3H, s, C₁₅-CH₃), 2.37 (1H, m, Val-(CH₃)₂CH), 2.40 (1H, m, Val-CH₃), 2.86 (3H, s, N-CH₃), 3.00 (1H, m, C₂-H), 3.15 (1H, m, C₂-H), 3.94 (1H, dd, J=10.4, 7.3 Hz, Gly-2CH₂), 4.03 (1H, dd, J=17.4, 4.6 Hz, Val- α -H), 4.52 (1H, d, J=11.3 Hz, Gly-CH₂), 4.89 (1H, s, C₁₂-H), 4.93 (1H, s, C₉-H), 5.21 (3H, m, C₅-H, C₁₂-CH₂), 5.32 (1H, m, Ala-α-H), 6.01 (1H, s, C₇-H), 6.31 (1H, d, J=10.1 Hz, Ala-NH), 7.54 (1H, br, Gly-NH), ¹³C NMR (CDCl₃) δ 12.9, 17.8, 18.3, 18.5, 19.3, 26.5, 29.1, 29.7, 30.9, 32.6, 40.6, 42.9, 43.2, 43.5, 65.9, 83.7, 105.3, 129.4, 130.5, 136.8, 141.3, 145.9, 166.5, 168.2, 171.6, 172.7. HRMS (EI) *m*/*z* Calcd for C₂₈H₄₅N₃O₅: 503.3359. Found: 503.3358.

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23. Although we attempted the macrocyclization at the other sites as shown in Fig. 4 below, the desired cyclized products could not be obtained.



Figure 4. Attempted macrocyclization.

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